



# UNIVERSIDAD AUTÓNOMA CHAPINGO

DEPARTAMENTO DE ENSEÑANZA, INVESTIGACIÓN Y SERVICIO EN ZOOTECNIA  
POSGRADO EN PRODUCCIÓN ANIMAL

## DIVERSIDAD GENÉTICA PARA CARACTERÍSTICAS DE IMPORTANCIA ECONÓMICA EN BOVINOS SUIZO EUROPEO

### TESIS

Que como requisito parcial  
para obtener el grado de:

**MAESTRO EN CIENCIAS EN INNOVACIÓN GANADERA**

Presenta:

**MITZILIN ZULEICA TRUJANO CHAVEZ**

Bajo la supervisión de: **Agustín Ruíz Flores, Dr.**  
y **Paulino Pérez Rodríguez, Dr.**



Chapingo, Estado de México, diciembre 2021



**DIVERSIDAD GENÉTICA PARA CARACTERÍSTICAS  
DE IMPORTANCIA ECONÓMICA EN  
BOVINOS SUIZO EUROPEO**

Tesis realizada por **MITZILIN ZULEICA TRUJANO CHAVEZ**, bajo la supervisión del Comité Asesor indicado, aprobada por el mismo y aceptada como requisito parcial para obtener el grado de:

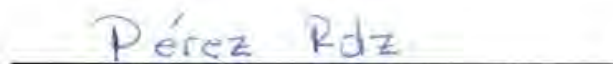
**MAESTRO EN CIENCIAS EN INNOVACIÓN GANADERA**

DIRECTOR:



Dr. Agustín Ruíz Flores

CODIRECTOR:



Dr. Paulino Pérez Rodríguez

ASESOR:



Dr. Luis Alberto Miranda Romero

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## **ABREVIATURAS**

|                 |                                                                                                         |
|-----------------|---------------------------------------------------------------------------------------------------------|
| ADN/DNA         | Ácido desoxirribonucleico<br>Deoxyribonucleic Acid                                                      |
| AFLP            | Polimorfismo en la longitud de fragmentos amplificados de DNA<br>Amplified Fragment Length Polymorphism |
| BC              | Corrección de Bonferroni<br>Bonferroni Correction                                                       |
| CAPS            | Secuencia polimórfica amplificada y cortada<br>Cleaved Amplified Polymorphic Sequences                  |
| CCR             | Tasa de concepción de la vaca<br>Cow Conception Rate                                                    |
| F <sub>IT</sub> | Coeficiente de consanguinidad<br>Inbreeding Coefficient                                                 |
| GS              | Selección genómica<br>Genomic Selection                                                                 |
| GWAS            | Estudio de asociación del genoma completo<br>Genome-Wide Association Study                              |
| HCR             | Tasa de concepción en vaquillas<br>Heifer Conception Rate                                               |

|      |                                                                                 |
|------|---------------------------------------------------------------------------------|
| He   | Heterocigosidad esperada<br>Expected heterocigosity                             |
| Ho   | Heterocigosidad observada<br>Observed heterocigosity                            |
| HWE  | Equilibrio Hardy-Weinberg<br>Hardy-Weinberg Equilibrium                         |
| MAS  | Selección asistida por marcadores<br>Marker Assisted Selection                  |
| NMDS | Escalamiento multidimensional no métrico<br>Non-metric Multidimensional Scaling |
| PAM  | Partición alrededor de medioides<br>Partition Around Mediods                    |
| PCA  | Análisis de componentes principales<br>Principal Component Analysis             |
| PL   | Vida productiva<br>Productive Life                                              |
| PR   | Tasa de preñez<br>Pregnancy Rate                                                |
| QTL  | Loci de características cuantitativas<br>Quantitative Trait Loci                |

|      |                                                                                                      |
|------|------------------------------------------------------------------------------------------------------|
| RAPD | Fragmentos polimórficos de ADN de amplificación aleatoria<br>Random Amplification of Polymorphic DNA |
| RFLP | Polimorfismo de longitud de fragmentos de restricción<br>Restriction Fragment Length Polymorphisms   |
| SNP  | Polimorfismo de nucleótido simple<br>Single Nucleotide Polymorphism                                  |
| SSR  | Repetición de secuencia simple o microsatélite<br>Simple sequence repeat                             |
| RBM  | Resistencia a mastitis bacteriana<br>Resistance to Bacterial Mastitis                                |
| RCM  | Resistencia a mastitis clínica<br>Resistance to Clinical Mastitis                                    |
| RM   | Resistencia a mastitis<br>Resistance to Mastitis                                                     |
| RSM  | Resistencia a mastitis subclínica<br>Resistance to Subclinical Mastitis                              |
| SM   | Susceptibilidad a mastitis<br>Susceptibility to Mastitis                                             |

## **DEDICATORIA**

A mis padres Enrique Trujano y Teresa Chávez por darme el regalo de la vida, enseñarme a dirigirme con rectitud y buenos valores; además, de brindarme su amor y apoyo siempre incondicionales.

A mi hermana y tía, Violeta Serrano y Marcela Chávez por su apoyo incondicional, amor, enseñanzas y consejos.

A mis sobrinos Naim y Raziel por enseñarme el significado del amor desinteresado y la protección.

A Reyna Sánchez, por ser un pilar indispensable en las nuevas etapas de mi vida, por su apoyo y amor incondicional.

A Diana Olivera, pese a la distancia, la mejor amiga que me ha acompañado en este y otros proyectos de mi vida.

## **AGRADECIMIENTOS**

A la Universidad Autónoma Chapingo, por acogerme y darme más de lo que podía pedir, por permitirme formar parte de su gran comunidad y por brindarme conocimiento.

Al Pueblo de México, que a través del Consejo Nacional de Ciencia y Tecnología (CONACyT), me brindó el apoyo económico para poder realizar mis estudios de Maestría y las presentes investigaciones.

Al Dr. Agustín Ruíz Flores por creer en mí, dándome su apoyo incondicional, supervisión, consejo y motivación en esta investigación y durante toda mi estancia en el Departamento de Zootecnia.

Al Dr. Paulino Pérez Rodríguez por su guía, apoyo, supervisión e indispensable contribución en los análisis estadísticos de esta investigación.

Al Dr. Luis Alberto Miranda Romero por su contribución en esta investigación.

## DATOS BIOGRÁFICOS

### Mitzilin Zuleica Trujano Chavez

Lugar y fecha de nacimiento: Texcoco, Estado de México; 28 de mayo de 1997.

CURP: TUCM970528MMCRHT07

Profesión: Ingeniero Agrónomo Especialista en Zootecnia

Cédula Profesional: 12203347



### Desarrollo Académico

2020-2021 Maestría en Ciencias en Innovación Ganadera. Universidad Autónoma Chapingo.

2012-2019 Ingeniero Agrónomo Especialista en Zootecnia. Universidad Autónoma Chapingo.

### Formación Adicional: Cursos

2021 Writing in the Sciences. Stanford University.

2021 The R Programming Environment. Johns Hopkins University.

2021 Matrix Methods. University of Minnesota.

2020 Cálculo Diferencial e Integral unidos por el Teorema Fundamental del Cálculo. Instituto Tecnológico y de Estudios Superiores de Monterrey (ITESM).

2019 Introducción a la Programación Estadística con R. Universidad Nacional Autónoma de México (UNAM).

## RESUMEN GENERAL

### DIVERSIDAD GENÉTICA PARA CARACTERÍSTICAS DE IMPORTANCIA ECONÓMICA EN BOVINOS SUIZO EUROPEO<sup>1</sup>

El objetivo de la presente investigación fue estudiar la diversidad genética de una población mexicana de bovinos Suizo Europeo para varias características de importancia económica, mediante parámetros poblacionales como heterocigosidad observada, coeficiente de consanguinidad y frecuencia de alelos favorables. Además, se usó estadística multivariada para visualizar la dispersión de los animales con base en su perfil genotípico (análisis de componentes principales, escalamiento multidimensional no métrico y agrupación por los métodos de agrupamiento jerárquico y K-means). Todos los análisis se realizaron con al menos 150 animales genotipados con el chip Genomic Profile Bovine LD de 30K y 50K marcadores de polimorfismo de nucleótido simple. En el primer estudio se encontró que, para muchos de los genes relacionados con calidad de la carne, la población mexicana de Suizo Europeo presentó alta frecuencia alélica favorable para genes asociados con mayor suavidad, mayor marmoleo, menor cantidad de ácidos grasos saturados, y menor fuerza de corte (CAPN1 y DGAT2). En el segundo estudio se midió la diversidad genética para características reproductivas, donde los genes CSNK1E, DNAH11, DSC2, IBSP y OCLN afectaron la mayoría de las características y fueron altamente informativos por lo que se encontraron al menos dos grupos con potencial de selección. En el tercer estudio se determinó la diversidad y estructura genética de la misma población para resistencia y susceptibilidad a mastitis. Si bien, 13.3% de los animales fueron altamente consanguíneos (>50%), hubo grupos altamente heterocigotos, indicativo de diversidad y por tanto favorables. Aunque, los genotipos homocigotos para genes inmunológicos (grupo CXCL) podrían ser favorables para características como resistencia a mastitis clínica y subclínica. En los tres estudios se encontró evidencia del potencial genético de la población mexicana de Suizo Europeo, para mejorar características complejas asociadas con calidad de la carne, reproducción y mastitis.

**Palabras clave:** consanguinidad, estadística multivariada, genes candidatos, SNP

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<sup>1</sup> Tesis de Maestría en Ciencias en Innovación Ganadera, Posgrado en Producción Animal, Universidad Autónoma Chapingo

Autor: Mitzilin Zuleica Trujano Chavez

Director: Agustín Ruíz Flores. Codirector: Paulino Pérez Rodríguez



## **GENERAL ABSTRACT**

### **GENETIC DIVERSITY FOR TRAITS OF ECONOMIC IMPORTANCE IN BRAUNVIEH CATTLE<sup>2</sup>**

The aim of this research was to unravel the genetic diversity of a Braunvieh cattle population, using population parameters such as observed heterozygosity, inbreeding coefficient, and favorable allele frequency. Additionally, multivariate statistics were used to visualize the dispersion of the animals based on their genotypic profile using principal component analysis, non-metric multidimensional scaling and grouping by the Brand and K-means methods. All the analyses were performed for at least 150 animals genotyped with the Genomic Profile Bovine LD chip of 30K and 50K single nucleotide polymorphism markers. In the first study it was found that, for many of the genes related to meat quality, the Mexican Braunvieh population presented a high favorable allelic frequency for genes associated with increased tenderness and marbling, less amount of saturated fatty acids, and less cutting force (CAPN1 and DGAT2). In the second study, genetic diversity was determined for reproductive traits, where the CSNK1E, DNAH11, DSC2, IBSP, and OCLN genes affected most of the traits and were highly informative; therefore, at least two groups with selection potential were found. Although 13.3% were highly inbred animals (>50%), there were highly heterozygous groups in terms of the traits studied, a favorable indicator of the presence of genetic diversity. Even though, the homozygous genotypes for immunologic genes (group CXCL) might be favorable for traits such as resistance to subclinical mastitis and resistance to clinical mastitis. In all the three studies, there is evidence of the genetic potential of the Mexican Braunvieh population, to improve complex traits associated with meat quality, reproduction, and mastitis.

**Key words:** Inbreeding, multivariate statistics, candidate genes, SNP

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<sup>2</sup> Master of Science Thesis in Livestock Innovation, Graduate Program in Animal Production, Universidad Autónoma Chapingo

Author: Mitzilin Zuleica Trujano Chavez

Advisor: Agustín Ruíz Flores. Co-advisor: Paulino Pérez Rodríguez

## **CAPÍTULO 1. INTRODUCCIÓN GENERAL**

Durante la historia de selección genética el principal factor para identificar una característica a mejorar ha sido su importancia para generar valor económico. Hasta hace tres décadas en la producción pecuaria se seleccionaban animales sólo para mejorar características productivas como producción de leche, carne, huevo, entre otras. En bovinos Holstein la intensa selección por producción de leche generó la disminución del comportamiento reproductivo de las vacas. Después de la década de 1970, se buscaron otras opciones para mejorar las estrategias de selección para lograr incrementar la ganancia genética de características en forma conjunta (Miglior et al., 2017; Veronese et al., 2019).

De acuerdo con Meuwissen, Hayes y Goddard (2016), algunos de los obstáculos con los que se enfrentó la selección genética en ganado bovino fueron las características cuantitativas con baja heredabilidad, que se miden tarde en la vida del animal y que son difíciles de medir -como las características reproductivas, de calidad de la carne y canal-; por tanto, el mejoramiento basado en fenotipos es un proceso tardado con poca ganancia genética anual.

Desde 1974 con el surgimiento de los primeros marcadores moleculares (Yang et al., 2013), empezó a desarrollarse una nueva estrategia para mejorar características complejas. El uso de los marcadores permitió el mejoramiento genético de animales basándose en su genotipo real, más que en la información fenotípica, lo que atendió en parte el problema planteado por Meuwissen et al. (2016). Además, la asociación de marcadores moleculares con características de importancia económica (donde se obtienen genes candidatos) permite desentrañar la base genética de los procesos fisiológicos relacionados con las características (Yan et al., 2013).

Actualmente, el genotipado de animales es un procedimiento sencillo, basta con tomar una muestra de pelo o sangre de los animales para obtener sus genotipos (Meuwissen et al., 2016). Los marcadores más utilizados para selección genómica son los polimorfismos de nucleótido simple (SNP), dado que su uso

generalizado ha reducido hasta un nivel rentable los costos de genotipado en sistemas comerciales y de investigación (Wiggans et al., 2017).

De acuerdo con Ali et al. (2019), el aprovechamiento de la variación genética es un elemento clave en el desarrollo y uso de cualquier raza en programas de mejoramiento. Para determinar los niveles de variación se han realizado múltiples estudios de diversidad y estructura genética usando la información de los SNP en ganado bovino. Estos estudios se desarrollaron principalmente para características complejas como reproducción, salud, reducción de emisiones de gases de efecto invernadero y resistencia a parásitos (Weller et al., 2017; Xia et al., 2018).

Las razas adaptadas a ambientes adversos poseen mayor variabilidad genética (Xia et al., 2018), debido a la necesidad del organismo para ser versátil ante los cambios del ambiente. En México, uno de los sistemas que afronta mayores adversidades climáticas para la producción bovina es el de doble propósito, ubicado en las regiones tropicales del país. Los bovinos adaptados a estos ambientes con resultados satisfactorios son *Bos indicus*, como la raza Indubrasil (Peixoto et al., 2021) o la Guzerat (Campos et al., 2017). Sin embargo, actualmente la raza Suizo Europeo ha tenido un auge importante en estos sistemas de producción debido a los buenos resultados en producción de carne como raza paterna (Rojo-Rubio, 2009).

La raza Suizo Europeo, a pesar de estar adaptada a climas templado-fríos (Bhati et al., 2020), se ha consolidado como una de las favoritas en los sistemas doble propósito como raza paterna (Rojo-Rubio, 2009). Por tanto, se esperaría que la población mexicana de Suizo Europeo en estudio tuviera una variabilidad genética importante, que pudiera aprovecharse en programas de mejoramiento.

Utilizando ejemplares de la raza Suizo Europeo y sus razas emparentadas, se hicieron estudios de diversidad genética para desentrañar la base genética de su adaptación a los ambientes. En el estudio de Moscarelli et al. (2020), se encontraron diferencias genéticas importantes entre varias razas derivadas de la

Braunvieh original, de acuerdo con el ambiente donde terminaron de adaptarse. Por otro lado, Bhati et al. (2020), en un estudio de diversidad con 49 toros ancestros, encontraron que la raza posee alta diversidad genética, frente a otras razas europeas de doble propósito.

### **Objetivo general**

Determinar la diversidad genética de una población de bovinos Suizo Europeo, mediante parámetros poblacionales como heterocigosidad observada, coeficiente de consanguinidad y frecuencia de alelos favorable; además, de usar estadística multivariada para visualizar la dispersión de los animales con base en su perfil genotípico.

### **Objetivos específicos**

1. Estimar las frecuencias génicas y genotípicas para loci de genes candidatos de calidad de la carne en una población mexicana de bovinos Suizo Europeo.
2. Caracterizar genéticamente una población mexicana de Suizo Europeo para vida productiva, tasa de preñez, y la tasa de concepción en vacas y vaquillas, mediante un estudio de diversidad genética enfocado en genotipos y alelos favorables para SNPs intragénicos ( $\pm 25K$  pb) de genes candidatos, usando los índices de diversidad de heterocigosidad esperada y observada, algoritmos de agrupamiento, análisis de escalamiento multidimensional no métrico y estimación de frecuencias génicas.
3. Caracterizar genéticamente a la población mexicana de Suizo Europeo para las características resistencia y susceptibilidad a mastitis, resistencia a mastitis bacteriana y mastitis subclínica, y mastitis clínica para SNPs intragénicos ( $\pm 25K$  pb) de genes candidatos, usando los índices de diversidad de heterocigosidad esperada y observada, coeficientes de consanguinidad, algoritmos de agrupamiento y análisis de componentes principales.

Este documento de tesis está compuesto por seis capítulos. El Capítulo 2 contiene la revisión de literatura sobre marcadores moleculares, estudios de asociación del genoma completo y el desarrollo de genes candidatos; además, se revisa de manera general los principales descubrimientos de genes candidatos para el grupo de características en estudio; este capítulo concluye con una breve descripción sobre los tipos de estudios de diversidad realizados en el mundo.

En el Capítulo 3 se describe la caracterización genética a través de las frecuencias de alelos favorables para Suizo Europeo. El Capítulo 4 caracteriza a esta población de Suizo Europeo para características reproductivas de acuerdo con sus alelos favorables, se reportan algunos genes con mayor oportunidad de selección por la diversidad de sus alelos.

El Capítulo 5 describe la diversidad y estructura genética de la población bovina estudiada para resistencia y susceptibilidad a mastitis, que a diferencia de los dos capítulos anteriores tiene un enfoque conservativo de versatilidad asociada con los genes más heterocigóticos. Finalmente, en el Capítulo 6 se presentan las conclusiones generales de los estudios que componen esta tesis (capítulos 3 a 5).

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## **CAPÍTULO 2. REVISIÓN DE LITERATURA**

### **2.1 Marcadores moleculares**

De acuerdo con Singh et al. (2014), un marcador genético es un término general aplicable a un fenotipo con base genética, que ayuda a evaluar la variabilidad fenotípica observada en los individuos. Los mismos autores señalan que existen diferentes tipos de marcadores genéticos que son base del mejoramiento genético animal, estos pueden ser morfológicos, productivos, bioquímicos y moleculares.

Un marcador molecular es producto del avance en metodologías moleculares para análisis genético, actualmente es lo más preciso para explicar los fenotipos de los animales (Marwal & Gaur, 2020). En concreto, un marcador molecular es una variación en un segmento de ADN -o gen con ubicación conocida- específica entre individuos asociada con características de importancia económica (Nadeem et al., 2017; Singh et al., 2014).

Singh et al. (2014), señalan que estas variaciones en el ADN pueden ser delecciones, inserciones, translocaciones, duplicaciones y mutaciones puntuales. En función de estas variaciones existen diferentes tipos de marcadores, cuyas propiedades varían; en general la historia de los marcadores se puede resumir en tres grupos, que se describen a continuación.

#### **2.1.1 Marcadores RFLP, RAPD y AFLP**

Según Yang, Kang, Yang & Fanq (2013), dentro de los marcadores obtenidos mediante hibridación, se encuentran los de polimorfismo de longitud de fragmentos de restricción (RFLP; por sus siglas en inglés), que fueron los primeros marcadores basados en ADN y se desarrollaron en 1974. De acuerdo con Singh et al. (2014), estos marcadores se basan en inserciones o delecciones en los sitios de restricción de las enzimas, donde se utilizan sondas para la identificación de los genes y la eventual genotipificación.



Las principales ventajas de los marcadores RFLP: son altamente reproducibles, codominantes, multialélicos y no requieren información de secuencia para generarse; mientras que, algunas de sus desventajas son: se requiere gran cantidad de ADN de alta calidad, sus procesos de obtención son tardados y difíciles de automatizar (Hashim & Al-Shuhaib, 2019; Yang et al., 2013).

Yang et al. (2013) mencionan que en 1990 se desarrollaron por primera vez los marcadores de fragmentos polimórficos de ADN amplificados aleatoriamente (RAPD; por sus siglas en inglés), que se basan en la amplificación del ADN mediante PCR, usando un primer único de 10 nucleótidos. Nadeem et al. (2017) señalan que, una ventaja importante que comparten estos marcadores con los RFLP es que no necesitan información de secuencias para su desarrollo; además, el proceso de obtención de los RPD no involucra la técnica de la hibridación, es decir, es rápido, simple y eficiente. Sin embargo, poseen una desventaja importante, es un marcador dominante, lo que limita su aplicación en el mejoramiento animal a características explicadas por esta acción génica.

Un tercer marcador llegó a minimizar las desventajas en la obtención y aplicación de los RFLP y RAPD. Los marcadores de polimorfismo en la longitud de fragmentos amplificados de ADN (AFLP; por sus siglas en inglés), desarrollados en 1993, son la combinación de polimorfismo en sitios de restricción e hibridación de *primers* arbitrarios, es decir, en los AFLP se combina el poder de los RFLP con la simplicidad en el proceso de obtención de los AFPL (Nadeem et al., 2017). A pesar de estas ventajas, los AFLP son marcadores dominantes lo que limita su uso generalizado (Nadeem et al., 2017).

Yang et al. (2013) mencionan que los AFLPs destacan por su estabilidad genética, su desarrollo rápido y barato; además, a pesar de ser estrictamente dominantes, los AFLPs pueden ser útiles en genética de poblaciones y la caracterización del genoma completo, para evaluar y caracterizar los recursos genéticos animales. A pesar de estas aplicaciones, los AFLPs actualmente han

caído en desuso debido al surgimiento de otros marcadores de ADN que brindan información genética más precisa y de forma eficaz.

### **2.1.2 Marcadores SSR o microsatélites**

Ante las limitantes de los marcadores ya mencionados, surgió un nuevo tipo de marcador, llamado microsatélite o repeticiones de secuencia simple (SSR; por sus siglas en inglés). Estos marcadores son repeticiones cortas en tándem conformadas por, repeticiones de 2 a 3 bases (dinucleótidos y trinucleótidos) comúnmente. Los SSR se caracterizan por ser altamente polimórficos (hasta más de 20 alelos), debido a la gran cantidad de repeticiones en el genoma (Nadeem et al., 2017).

Los SSR poseen las mismas ventajas que los RFLP, tienen mayor repetibilidad y estabilidad que los RAPD, además de ser marcadores codominantes. Algunas de sus desventajas son: alto índice de error en el genotipado ante mutaciones no identificadas y desarrollo lento y costoso. Los microsatélites fueron los marcadores más utilizados durante años, hasta la llegada de los polimorfismos de nucleótido simple (SNP; por sus siglas en inglés) (Nadeem et al., 2017; Yang et al., 2013).

### **2.1.3 Marcadores SNP**

Los marcadores SNP son lo último en precisión a nivel de ADN, que representan el cambio de un solo par de bases presentes en la secuencia del genoma de un individuo; desarrollados por primera vez en 1996, son la unidad más pequeña de herencia, y su variación se puede deber a transiciones o transversiones sobre la base de sustitución de nucleótidos. Los métodos de genotipificación SNP más conocidos son: mediante desarrollo de RFLP (SNP-RFLP) y la técnica del marcador de secuencia polimórfica amplificada y cortada (CAPS; por sus siglas en inglés). El método consiste en secuenciar a los animales y encontrar diferencias en un solo par de bases entre ellos (Marwal & Gaur, 2020; Nadeem et al., 2017, Yang et al., 2013;).

De acuerdo con Yang et al. (2013), los marcadores SNP actualmente son de los marcadores más utilizados en mejoramiento genético, dado que son abundantes en el genoma, son genéticamente estables y cuyo análisis puede ser completamente automatizado, por ende, su desarrollo es de alto rendimiento; además, son marcadores codominantes por lo que es sencillo identificar animales heterocigóticos y homocigóticos.

## **2.2 Estudios de asociación del genoma completo**

De acuerdo con Tian et al. (2020), el estudio de asociación del genoma completo (GWAS; por sus siglas en inglés) es una técnica genética clave para explorar la base genética de características de interés en los animales. Estos mismos autores señalan que GWAS pretende detectar la asociación de variaciones genéticas específicas con un fenotipo, el método implica el análisis de genomas de muchos individuos para realizar la búsqueda de marcadores moleculares (sobre todo SNPs), útiles para predecir un valor fenotípico asociado con una característica en estudio.

### **2.2.1 Metodología básica de GWAS**

Uitterlinden (2016) describe un procedimiento básico para realizar un GWAS, el cual involucra los siguientes pasos:

1. *Colección del ADN/genotipado*: este autor señala que actualmente existen muchas opciones para realizar el genotipado, en todas las especies y con varios tipos de muestra disponibles, usualmente se recurre a las muestras de pelo y sangre. Existen diversos chips para obtener los genotipos, entre los que destacan Affymetrix e Illumina. Para encontrar resultados reveladores en un GWAS es clave tener genotipos de dos grupos de individuos: un grupo control y un grupo con la característica deseada.
2. *Imputación de genotipos*: en muchas ocasiones la información genómica resultante de la amplificación de muestras de ADN no es

concluyente, lo que sugiere el desconocimiento de los genotipos para ciertos marcadores en algunos individuos. Imputar los genotipos faltantes es una alternativa viable para mejorar la precisión de GWAS (Uitterlinden, 2016); de acuerdo con Browning et al. (2018), Beagle 5.2 es uno de los programas más usados para imputar genotipos.

3. *Análisis de datos:* hasta este punto, se tienen dos bases de datos, la primera, contiene la información genotípica de los individuos; mientras que, la segunda, contiene el valor fenotípico de la característica de interés para esos mismos individuos. La información resultante del genotipado debe recodificarse para pasar de una escala categórica (AA, AB, BB) a valores numéricos (0, 1 y 2) en la que se cuenta la presencia de cierto alelo.

De acuerdo con la naturaleza de los fenotipos, es posible realizar regresión logística (valores dicotómicos) o regresión lineal (valores continuos); cualquier programa de análisis estadístico es útil mientras pueda procesar grandes matrices. Típicamente hay dos formas de representar los resultados de un GWAS: representados como valores de  $p$  para la prueba de asociación (observados vs. esperados) en una gráfica cuantil-cuantil o como una gráfica de Manhattan (Uitterlinden, 2016).

Un último paso no enumerado por Uitterlinden (2016), es la caracterización del SNP significativo para la característica. Una vez que se ha identificado que un SNP posee un efecto sobre la característica estudiada, se procede a estudiar este segmento de ADN. Recordando que ese marcador específicamente puede ser la causa de variación fenotípica, o bien algún otro segmento de ADN localizado cerca de él, gracias al desequilibrio de ligamiento. Este último paso requiere de múltiples estudios, debido a que es posible que el segmento tenga carácter cuantitativo, lo que involucra mayor trabajo de laboratorio para encontrar los procedimientos fisiológicos que le dan lugar.

### 2.2.2 Métodos estadísticos para GWAS

En el subtema anterior se comentó acerca del análisis estadístico para realizar GWAS. La regresión lineal simple es el método más elemental para realizar la asociación de los marcadores con los fenotipos. Al respecto Hayes (2013), describe el modelo estadístico para la regresión de marcador único, cuyo uso se fundamenta en la explotación del desequilibrio de ligamiento.

Hayes (2013) señala que, el desequilibrio de ligamiento se refiere a las asociaciones en una población entre marcadores moleculares y mutaciones causales o loci de características cuantitativas (QTL; por sus siglas en inglés). Tales asociaciones surgen debido a que algunos segmentos del genoma -provenientes de un mismo cromosoma- se heredan juntos a través de las generaciones, es decir no hay efectos recombinantes. A estos segmentos de cromosoma se les conoce como haplotipos, usados en el modelo de GWAS más sencillo: regresión marcador por marcador.

Esta regresión, que estudia la asociación entre marcadores moleculares y fenotipos, está dada por (Hayes, 2013):

$$\mathbf{y} = \mathbf{W}\mathbf{b} + \mathbf{X}\mathbf{g} + \mathbf{e},$$

donde  $\mathbf{y}$  es el vector de fenotipos,  $\mathbf{W}$  es la matriz de diseño que relaciona los datos con los efectos fijos fenotípicos,  $\mathbf{b}$  es un vector de efectos fijos como la media, efectos de estructura poblacional y edad,  $\mathbf{X}$  es el vector que conecta los datos con los efectos de marcador,  $\mathbf{g}$  es el efecto del marcador y  $\mathbf{e}$  es el vector de desviaciones aleatorias con el supuesto  $e_i \sim N(0, \sigma_e^2)$ , donde  $\sigma_e^2$  es la varianza del error.

La hipótesis nula para el modelo establece que el marcador no tiene efecto en la característica; mientras que, la hipótesis alterna establece un efecto significativo sobre la característica, lo anterior se afirma debido a la existencia de desequilibrio de ligamiento entre los marcadores y un posible QTL (Hayes, 2013).

El modelo anterior es un análisis de un solo SNP por vez, en consecuencia, el enfoque de GWAS generalmente usa SNPs individuales para evaluar su asociación con un fenotipo de interés mientras ignora otros SNPs. Sin embargo, la mayoría de las características de interés en animales son cuantitativas, es decir, una parte no conocida de la heredabilidad incluye efectos epistáticos además de aditivos. Otra limitante del uso de este modelo GWAS es la gran capacidad computacional necesaria para llevar a cabo las operaciones matriciales del modelo mixto (Mortezaei & Tavallaee, 2021).

Para solucionar estos problemas se han desarrollado metodologías que permiten dar un enfoque multi-loci a GWAS y que ayudan a minimizar el problema de grandes bases de datos en la obtención de los resultados. Al respecto, Mortezaei y Tavallaee (2021) enlistan varios tipos de procedimientos alternativos para la realización de GWAS, dos de los cuales se muestran a continuación.

#### **2.2.2.1 Aprendizaje automático**

Usando el aprendizaje automático es posible descubrir patrones ocultos dentro de la base de datos de los marcadores, que pueden ayudar a desentrañar la base genética completa de las características. El aprendizaje automático se basa en la inteligencia artificial, donde las computadoras pueden tomar decisiones aprendiendo de los datos. La regresión de marcador único es una forma incipiente del aprendizaje automático; mientras que bosques aleatorios o metodologías basadas en el principio de máxima verosimilitud son herramientas nuevas con aplicaciones novedosas. Las máquinas de soporte vectorial son modelos que pueden usarse en conjunto con la regresión simple usada en marcador único (Díez et al., 2021; Mortezaei & Tavallaee, 2021).

#### **2.2.2.2 Análisis de asociación retrospectivo**

Considerando los múltiples efectos no genéticos asociados con un fenotipo, se han creado modelos de asociación que incluyen covariables para corregir posibles variaciones no controladas en el ambiente, a estos procedimientos se

les conoce como análisis de asociación retrospectivo, que prometen ser más precisos que los modelos comunes prospectivos para variables binarias; sin embargo, esto no se ha comprobado. Dos de las metodologías estadísticas para realizar este tipo de análisis son la prueba de características binarias longitudinales basada en ecuaciones y la prueba de asociación basada en modelos mixtos lineales generalizados retrospectivos (Mortezaei & Tavallaee, 2021).

### **2.2.3 Genes candidatos**

Derivado de GWAS, los genes candidatos son aquellos que se asocian con una característica. Los genes son segmentos de ADN con ubicación fija en el genoma, se pueden ubicar mediante el número de cromosoma y su rango de posición en pares de bases. En los estudios de asociación, los marcadores usados se encuentran dispersos casi siempre de forma aleatoria a lo largo del genoma, cuando se encuentra un marcador significativo para una característica, se recupera la posición de este marcador  $\pm 25,000$  pares de bases y entonces se asigna a un gen que lo contenga. Este gen que contiene el marcador significativo se conoce como gen candidato. Los genes candidatos contienen por lo menos un segmento de ADN que afecta una característica, gracias al desequilibrio de ligamiento (Mota et al., 2017a).

Es más efectivo estudiar genes candidatos que marcadores por sí solos. Existen genes que codifican productos funcionales que tienen efecto sobre diferentes características en el organismo (Gerstein et al., 2007). Por ejemplo, el gen CAPN1, es un gen candidato en bovinos para suavidad y marmoleo de la carne (Page et al., 2004). Fisiológicamente podrían existir hasta dos explicaciones diferentes sobre el efecto del gen y seguramente estas características poseen una correlación positiva dado que los segmentos de ADN que las define están en el mismo gen y poseen equilibrio de ligamiento.

## **2.3 Genes candidatos en bovinos**

Muchos de los genes que se abordarán a continuación, no fueron incluidos en los estudios de los capítulos 3 a 5, debido a que no se encontraban sus posiciones en la base de datos de la que se dispuso para el estudio. Con ello se trata de presentar una perspectiva más completa de lo encontrado para *Bos taurus* referente a los genes candidatos más importantes por característica.

### **2.3.1 Calidad de la carne**

Page et al. (2004) y White et al. (2005), identificaron en el ganado *Bos taurus* y *Bos indicus* SNP en la posición del gen CAPN que se asocian con suavidad de la carne, y estos SNP se les llamó CAPN1-316 y CAPN1-4751. Por su parte, Soria et al. (2010) reportaron otro SNP, asociado al mismo fenotipo, el SNP CAPN1-530.

El alelo C, tanto en CAPN1-316 (Page et al., 2004) como en CAPN1-4751, se ha asociado con valores favorables de terneza de la carne. Sin embargo, este alelo se ha encontrado en baja proporción en algunas poblaciones (Casas et al., 2006; Page et al., 2004; White et al., 2005).

Para el gen CAST se identificó un SNP de sustitución de citosina por timina que está asociado con la terneza de la carne. Los animales homocigotos para timina producen carne con menores valores de fuerza de corte que aquellos homocigotos para citosina (Casas et al., 2006).

El sistema CAPN/CAST está involucrado en la miogénesis y la comunicación entre células, la diferenciación celular y la degradación de proteínas musculares y de proteínas de la membrana celular durante la fusión de los mioblastos, al formarse el miotubo, y de células satélites (Moyen et al., 2004, Barnoy et al., 2005). Según lo reportado por Casas et al. (2006), los polimorfismos relacionados con este sistema pueden influir en el índice de producción y suavidad de la carne.



Por su parte, el gen DGAT1 codifica la proteína acetil Coenzima A diacilglicerol aciltransferasa (DGAT) que cataliza el paso final en la síntesis de triacilglicerol (Casos et al., 1998; Grisart et al., 2002). De acuerdo con Casos et al. (1998), esta enzima también puede estar involucrada en la síntesis y almacenamiento de lípidos, y dos marcadores de este gen se han asociado con la variación en el contenido de grasa de la leche y la carne. Al respecto, Curi et al. (2011) encontraron que el gen DGAT1 es polimórfico en ganado Nelore y que tiene una posible asociación con la cantidad de grasa de la carne.

Los polimorfismos del gen TCAP se encuentran entre los factores genéticos importantes involucrados en la calidad de la canal en el ganado vacuno; el gen titin-cap codifica telethonin, una proteína que interactúa con la titina en la línea Z; las asociaciones entre los SNP del gen TCAP y el veteado en *Bos taurus* mostraron que la sustitución de guanina por adenina redujo la puntuación de veteado (Cheong et al., 2007).

La familia MYOD, conformada por los genes MYOD1 o MYF3, MYOG, MYF4 (miogenina), MYF5 y MYF6 (herculina), juegan un papel clave en el crecimiento y el desarrollo muscular, se consideran genes candidatos para las características de producción de carne en animales (Bhuiyan et al., 2009). La familia está involucrada en la regulación de la hipertrofia muscular y la hiperplasia (Bhuiyan et al., 2009; Houba & Te Pas, 2004).

Según Houba y Te Pas, (2004), el gen MYOG se expresa antes del nacimiento; mientras que, Te Pas y Soumilion (2001) afirman que, en el período posnatal, puede asociarse con la reparación del daño en las fibras musculares y el crecimiento hipertrófico.

En cerdos, MYOG se ha asociado con un aumento en el número y el área transversal de las fibras musculares (Kim et al., 2009), el peso al nacer y en canal, y la tasa de crecimiento (Te Pas et al., 1999). Sin embargo, en el ganado *Bos taurus*, el efecto se observó solo en la capacidad de retención de agua y la suavidad de la carne (Ujan et al., 2011).

Las anquirinas son proteínas involucradas en las interacciones entre la membrana plasmática y el citoesqueleto (Bagnato et al., 2003). Según Rubtsov y Lopina (2000), los genes ANK1, ANK2 y ANK3 codifican anquirinas expresadas en tejidos específicos. El empalme alternativo de ANK1 produce ankyrin que tiene un dominio sensible a la proteólisis y puede ser el objetivo de la calpaína, lo que afecta la suavidad de la carne en bovinos.

De acuerdo con Curi et al. (2011), el gen FABP4 codifica la proteína de unión a ácidos grasos que se encuentra en los adipocitos, sus funciones incluyen la absorción, el transporte y el metabolismo de los ácidos grasos. Curi et al. (2011) reportaron que en ganado Nelore puro el gen puede tener un efecto notorio; aunque el I polimorfismo FABP4/Msp A1I no mostró asociación con las características de área del ojo de la costilla, cantidad de grasa, grasa intramuscular y fuerza de corte e índice de fragmentación miofibrilar en animales cruzados.

### **2.3.2 Características reproductivas**

Entre las características reproductivas de mayor importancia en bovinos están: tasa de concepción para vaquillas y vacas adultas, vida productiva y mérito neto global (Cochran, Cole, Null & Hansen, 2013). Este tipo de características poseen baja heredabilidad y toma mucho tiempo medirlas durante la vida del individuo, la precisión en la predicción también es baja, por lo que encontrar genes candidatos fue la primera opción para mejorar las predicciones y el mejoramiento genético, sobre todo en la raza Holstein (Cochran et al., 2013).

En estudios donde se utiliza GWAS, se han probado miles de genes candidatos reportados en literatura, encontrando de 20 a 50 genes candidatos relacionados de forma importante con características reproductivas (Cochran et al., 2013; Pimentel et al., 2011).

Para tasa de concepción, Ortega et al. (2016) evaluaron diferentes marcadores SNP de genes candidatos en una población de vacas Holstein. Cerca de 57% de

los SNP tuvieron un efecto importante en la característica. Los genes candidatos con efecto significativo fueron: COQ9, EPAS1, CAST, C7H19orf60 y MRPL48. El gen CAST se había discutido en la sección de genes candidatos para calidad de la carne, lo que refuerza la idea antes planteada de que los genes candidatos son un factor de estudio importante, que pueden afectar varias características cuantitativas.

Para la característica de tasa de concepción en vacas, en el mismo estudio de Ortega et al. (2016), se confirmó un efecto importante de los genes candidatos GOLGA4, COQ9, EPAS1 y MRLP48 en la característica. Estos mismos autores reportan genes candidatos para la tasa de concepción en vaquillas HSD17B7, AP3B1, HSD17B12 y CACNA1D. Algunos de los genes reportados asociados con fertilidad en general, están involucrados en la esteroidogénesis o están regulados por esteroides, lo que explica que algunos de estos genes coincidan con genes candidatos asociados con características sobre contenido de grasa (Ortega et al., 2016).

En ganado Canchim -raza compuesta por 62.5% Charolais y 37.5% Cebú- Buzanskas et al. (2017) encontraron que el gen RAP1B participa en varias vías que afectan el comportamiento reproductivo, la principal vía es la del receptor de la hormona liberadora de gonadotropinas. Por otro lado, los mismos autores señalan que MED30 y TRHR también son genes candidatos importantes relacionados con la hormona tiroidea que afecta directamente la reproducción.

Buzanskas et al. (2017), al igual que Ortega et al. (2016), hallaron que los genes FABP5, FABP12, PEX2 y MED30 están involucrados en el metabolismo de ácidos grasos, colesterol y triacilglicerol para la producción de hormonas sexuales, sobre todo para la producción de testosterona.

Según Buzanskas et al. (2017), el gen que afecta significativamente el desempeño reproductivo de machos Canchim es UBQLN3, que se expresa en los testículos y actúa en la espermatogénesis. De acuerdo con estos autores,

este gen está involucrado en el procesamiento de proteínas en la vía del retículo endoplásmico.

### **2.3.3 Resistencia y susceptibilidad a mastitis**

En ganado bovino la mastitis es una enfermedad importante de la glándula mamaria, que causa pérdidas económicas y cuya base genética empezó a estudiarse hace relativamente poco. Algunos estudios han revelado genes candidatos (GC, NPFFR2, TRAPPC, ARHGAP39, LY6K, LY6D, LYNX1, LYPD2, SLURP1 y PSCA) asociados con el conteo de células somáticas para ganado Holstein (Cai, Guldbrandsen, Lund, & Sahana, 2018; Moretti et al., 2021).

Moretti et al. (2021), encontraron genes candidatos asociados con el conteo en células somáticas en ganado Holstein. Los SNP se identificaron en regiones intragénicas de genes relacionados con componentes clave del sistema inmunológico: CXCR1, DCK, NOD2, MBL2, MBL1 y M-SAA3.2. Por ejemplo, los receptores de quimiocinas CXCR 1 y 2 son genes parálogos que codifican los principales receptores de citocinas proinflamatorias (Moretti et al., 2021), que impactan directamente el proceso de inflamación de la glándula mamaria.

Kurz et al. (2019), encontraron grupos de animales Holstein resistentes y susceptibles a mastitis clínica midiendo la longitud del pezón como variable de respuesta. Encontrando que 7.6% de las vacas presentaban un genotipo para resistencia y 12.5% presentaron susceptibilidad genética para la enfermedad. Estos mismos autores encontraron que el gen mayormente asociado con la longitud del pezón, RASGRP1, es un gen candidato de nuevo descubrimiento que podría ser clave para seleccionar animales con genotipos favorables para disminuir la tasa de incidencia de mastitis en los hatos.

La mastitis es una enfermedad multifactorial, y por ende difícil de desentrañar genéticamente. Algunos enfoques que se han tomado para atender esta problemática se relacionan con la resistencia a bacterias específicas que promueven la aparición de la mastitis. Por ejemplo, Liabin, Chen y Chen (2019)

identificaron genes candidatos asociados con mastitis provocada por *Escherichia coli*, encontrando genes distintos a los que regularmente se hallan en los estudios de GWAS para resistencia en general. Los genes candidatos encontrados fueron IL8RB, CXL6y y MMP9.

Un estudio de Song et al. (2016) reveló genes candidatos asociados con la susceptibilidad a mastitis subclínica provocada por *Staphylococcus aureus* en ganado Holstein. Se encontró que los genes NRG1, MST1 y NAT9 se correlacionaron de manera importante con la característica.

### **2.3.4 Resistencia a garrapatas**

Otra característica difícil de mejorar mediante mejoramiento genético convencional es la resistencia y susceptibilidad a garrapatas. Es difícil de medir y su heredabilidad es baja. Con las metodologías actuales de asociación del genoma e identificación de genes candidatos, el mejoramiento de este tipo de características se hace cada vez más factible. Al igual que en el caso de mastitis, los enfoques se han hecho más específicos para poder dar solución a los diferentes impactos de las garrapatas (Moré et al., 2019).

Los genes asociados con resistencia a garrapatas hallados por Moré et al. (2019) en un estudio con bovinos Braford, tienen influencia en el sistema inmunológico en los animales. En particular, los autores mencionan una vía de señalización de *Wingless* como un mecanismo candidato a resistencia, donde está involucrado el gen WNT7A y los genes IL6 e IL22 como promotores.

En el estudio de Mota et al. (2017b) se encontraron genes candidatos asociados con la resistencia y susceptibilidad a garrapatas en ganado Hereford y Braford, algunos de los genes asociados con resistencia más importantes fueron: CCL20, MYO6, BSP1, BSP3 y PLAUR; mientras que, los asociados con susceptibilidad fueron DNAH8, BOLA-DQA2 y SALL4.

## **2.4 Uso de los marcadores moleculares en mejoramiento genético animal**

### **2.4.1 Selección asistida por marcadores**

Los marcadores moleculares con el surgimiento de la selección asistida por marcadores (MAS; por sus siglas en inglés), prometieron en sus inicios ayudar de forma efectiva y rápida al mejoramiento genético para características importantes económicamente en animales y plantas (Jena & Mackill, 2008). De acuerdo con Jena y Mackill (2008), el propósito en MAS es monitorear la presencia o ausencia de genes de interés en poblaciones; por otro lado, Collard y Mackill (2008) definieron MAS como el proceso de usar marcadores de ADN para asistir en la selección de material para el mejoramiento genético.

En párrafos anteriores se discutieron algunos elementos esenciales para realizar MAS: el surgimiento de los marcadores moleculares, el mapeo de QTL y la identificación de genes candidatos. MAS es una metodología más eficiente que la evaluación fenotípica convencional, que conlleva un ahorro de tiempo, esfuerzo, recursos y dinero (Platten, Cobb & Zantua, 2019).

El primer elemento es el tipo de marcador a utilizar, actualmente el uso de SSR y SNP es lo más pertinente debido a que poseen características deseables para MAS, son confiables, necesitan poca cantidad y calidad de ADN, su procedimiento técnico es sencillo, poseen un alto nivel de polimorfismo y su costo los vuelve rentables (Collard & Mackill, 2008).

Otro elemento importante para el éxito de MAS es el mapeo de QTLs. Este procedimiento se realiza gracias al mapeo de ligamiento que se basa en la recombinación ocurrida durante la meiosis. Un mapeo de QTLs es un procedimiento que permite encontrar loci asociados con características de interés; por lo general este mapeo no es suficiente para designar a un segmento de ADN candidato definitivo, sino que se necesita realizar mapeo fino para ajustarse a un segmento de ADN aún más pequeño (Collard & Mackill, 2008; Platten et al., 2019).

Collard & Mackill (2008) describieron un flujo general para concretar el proceso de MAS:

1. *Desarrollo de las poblaciones*: selección parental e hibridación. Los estudios de MAS pretenden obtener la mayor cantidad de individuos heterocigóticos por lo que se busca obtenerlos mediante selección y cruce.
2. *Mapeo de QTLs*: construcción del mapa de ligamiento, evaluación fenotípica para la característica y análisis de QTL. Con este paso se desea obtener un primer acercamiento de los efectos del QTL con la característica.
3. *Validación de QTLs*: confirmación de la posición y efecto de los QTL, mapeo fino. En este paso se obtiene de forma más precisa la longitud y posición de los QTLs con efectos sobre la característica. Esta validación permite obtener resultados similares entre poblaciones similares.
4. *Validación de marcadores*: aprovechando el desequilibrio de ligamiento y teniendo las posiciones de los QTLs, es posible obtener marcadores que estén asociados con estos QTL y por tanto, tengan un efecto en la característica.
5. *Selección asistida por marcadores*: la información de marcadores es útil en la selección de individuos con genotipos favorables.

Actualmente MAS puede ser una metodología enriquecida haciendo uso de los genes candidatos, es decir de GWAS. Para características difíciles de mejorar mediante el método convencional, como las que se discutieron previamente, MAS puede ser de ayuda para seleccionar individuos con genotipos favorables en etapas tempranas, logrando ahorrar tiempo y dinero en el mejoramiento genético. MAS es una alternativa viable en el mejoramiento de plantas y animales (Collard & Mackill, 2008; Platten et al., 2019). Sin embargo, MAS tiene una desventaja importante, muchas de las características que trata de explicar son cuantitativas, es decir, están influenciadas por muchos genes, lo más grave es que un solo gen no llega a explicar más de 10% de la variación de una característica (Collard & Mackill, 2008).

### **2.4.2 Selección genómica**

La selección genómica (GS; por sus siglas en inglés) se refiere al uso de marcadores que cubren el genoma de los animales para predecir el valor como reproductores de los candidatos a selección de una característica cuantitativa, asumiendo que todos los QTL están en desequilibrio de ligamiento con al menos un marcador. La GS es posible gracias al gran número de SNPs que cubren los genomas de especies domésticas de interés, y los métodos existentes para determinar los genotipos eficientemente en un número grande de marcadores (Goddard, 2009; Goddard & Hayes, 2007).

Como una solución a las limitantes de MAS, la GS permite el mejoramiento de los animales usando toda la información disponible del genotipado. Para incluir la información de algunos marcadores en las predicciones, es posible localizar mutaciones causantes subyacentes a la variación genética o mapeo de QTLs. En la práctica estos programas de mejoramiento genético han tenido un impacto menor al esperado, lo que se atribuye a que la proporción de la varianza contabilizada por el QTL mapeado generalmente es pequeña y que los recursos financieros para realizar los mapeos son elevados, lo que limita el uso de MAS (Berry et al., 2016; de los Campos et al., 2013).

En GS se asume que todos los marcadores están unidos a un gen con efectos para una característica, concentrándose en la estimación de estos efectos más que en probar su significancia. La mayoría de las características se ven afectadas por un gran número de genes con efectos pequeños; la predicción de características complejas requiere considerar un gran número de variantes al mismo tiempo (de los Campos et al., 2013; Meuwissen et al., 2016).

La selección genómica según Meuwissen et al. (2016) se hace en dos etapas, la primera incluye el genotipado y fenotipado de animales de la población de entrenamiento para una determinada característica con el fin de estimar los efectos de los SNP. En la segunda etapa se genotipa a los individuos candidatos a selección, población de evaluación, al combinar la información obtenida en la



primera etapa (efectos SNP) y los genotipos se obtienen los valores genómicos que servirán para seleccionar los mejores individuos para la característica evaluada.

Las principales ventajas de GS son la reducción del intervalo entre generaciones, el incremento de la tasa de progreso genético, mayor exactitud en la predicción de valores genéticos, mejora en la exactitud de valores genómicos de animales jóvenes, especialmente para características de heredabilidad baja, limitadas por el sexo, medidas tarde en la vida de los animales, costosas de medir, o que requieren retar a los animales (Garrick, 2011; Garrick & Saatchi, 2011; Yin & König, 2016).

Según Meuwissen et al. (2016), existen tres condiciones que han permitido el amplio uso de información del ADN en las evaluaciones genómicas: 1) la metodología para realizar GS (Meuwissen et al., 2001), 2) la identificación de miles de marcadores SNP, y 3) las tecnologías de costo bajo para determinar eficientemente genotipos.

## **2.5 Estudios de diversidad genética en bovinos**

La diversidad genética se refiere a los diferentes polimorfismos presentes en los genes de individuos de una misma especie, o incluso de una raza o una población. En la actualidad los estudios de diversidad genética se han generalizado debido al surgimiento de marcadores moleculares de bajo costo y con buena distribución en el genoma para especies domésticas (SSR y SNP) (Doublet et al., 2019).

Existen dos enfoques principales para el estudio de la diversidad genética usando marcadores moleculares en bovinos. Un enfoque se relaciona con la consanguinidad y estructura de la población, los estudios se enfocan en observar cómo ha cambiado el coeficiente de consanguinidad en una población a lo largo de los años, o después de implementar un programa de mejoramiento (Doublet et al., 2019; Lozada-Soto et al., 2021). Por otro lado, existe el enfoque

conservacionista que pretende investigar la diversidad genética de razas relacionadas genéticamente cuyos ancestros comunes datan de hace cientos o miles de años, lo que pretende desentrañar la base genética de su adaptación a diferentes regiones (Bhati, Kadri, Crysanto, & Pausch, 2020; Freitas et al., 2021; Moscarelli et al., 2020).

En el caso de la presente serie de investigaciones, los parámetros relacionados con diversidad y estructura genética se utilizaron para observar la tendencia de los genotipos de marcadores SNP para genes candidatos asociados con características de importancia económica, encontrando valores para heterocigosidad observada ( $H_o$ ) similares a los de animales adaptados al trópico, aun cuando Suizo Europeo es una raza *Bos taurus* (Trujano-Chavez et al., 2021). Este enfoque es similar al utilizado por Freitas et al. (2021), quienes estudiaron la diversidad genética de razas adaptadas a diferentes ambientes para la característica de resistencia a estrés calórico.

La diversidad genética por lo regular se auxilia de parámetros poblacionales y visuales para poder ser caracterizada. Depende del enfoque que se desee tomar, es el tipo de parámetros a utilizar y las herramientas visuales propias de la estadística multivariada pertinentes. Generalmente, el coeficiente de consanguinidad y la  $H_o$  son dos parámetros que no pueden faltar en un estudio de diversidad genética; mientras que, el análisis de componentes principales (PCA, por sus siglas en inglés) y los árboles filogenéticos son los principales recursos gráficos usados para visualizar las relaciones genéticas (Bhati et al., 2020; Freitas et al., 2021).

### **2.5.1 Estadísticas básicas de la población**

En un estudio de diversidad genética (enfoque de conservación) con 10 razas bovinas y usando microsatélites, Agung et al. (2019) usaron los siguientes parámetros poblacionales: número observado de alelos, número efectivo de alelos,  $H_o$ , heterocigosidad esperada ( $H_e$ ), equilibrio Hardy-Weinberg. El número observado y efectivo de alelos son parámetros recurrentes en los estudios que

usan microsatélites, los cuáles poseen altos grados de polimorfismos, por lo que el conteo de alelos es necesario para describir la diversidad genética. Por otro lado,  $H_o$ ,  $H_e$  y el equilibrio Hardy-Weinberg son parámetros necesarios para cualquier tipo de estudio de diversidad, inclusive es posible calcularlos sin poseer información de marcadores moleculares.

Un enfoque similar al del párrafo anterior, fue propuesto por Freitas et al. (2021), quienes usaron la  $H_o$ ,  $H_e$  y el equilibrio Hardy-Weinberg como medidas de la diversidad genética en ganado europeo usando SNPs. Usaron otros parámetros interesantes como la distancia genética media por pares, el desequilibrio de ligamiento y el tamaño efectivo de la población. Lo anterior, para caracterizar la diversidad genética para el estrés térmico en tres razas.

Lozada-Soto et al. (2021) usando un enfoque de evaluación de estructura poblacional, caracterizaron la diversidad genética de ganado Angus usando los coeficientes de consanguinidad basados en pedigrí y en marcadores SNP; además, midieron los intervalos generacionales, el tamaño efectivo de población y el número efectivo de segmentos cromosómicos segregantes independientes. Estos autores se focalizaron en determinar los efectos de la GS sobre la diversidad genética de bovinos Angus, basándose en los coeficientes de consanguinidad más que en la frecuencia en heterocigosidad en los individuos.

En contraste, Xia et al. (2019), usaron el genoma mitocondrial para caracterizar a la raza Yunling, y los parámetros que utilizaron fueron menos convencionales que los de estudios mencionados en párrafos anteriores. Crearon haplotipos a partir de la secuenciación del genoma mitocondrial, para poder medir el número de haplotipos y los sitios variables, la diversidad de nucleótidos y el número medio de diferencias de nucleótidos.

### **2.5.2 Programas y estadística multivariable**

En la red existen una diversidad de *software* para calcular de forma sencilla los principales parámetros poblacionales usados en genética. Estos programas

también ofrecen herramientas estadísticas visuales para facilitar la interpretación de las relaciones genéticas entre los individuos en estudio.

El Boom de R (R Core Team, 2021) ha hecho que muchos paquetes incluyan en sus análisis herramientas para estudiar la diversidad genética. Uno de estos programas es *adegenet*, que ofrece el cálculo de las estadísticas poblacionales más sencillas como coeficientes de consanguinidad,  $H_o$ ,  $H_e$  y el equilibrio Hardy-Weinberg. Es versátil y ofrece estos parámetros para estudios que usan tanto SNPs como SSRs. Para estos últimos también calcula el número observado y efectivo de alelos (Jombart, 2008).

El paquete *adegenet* en principio fue creado para el análisis multivariado de marcadores, por lo que una de sus principales fortalezas es proporcionar herramientas visuales para la estructura y diversidad de la población. Lo hace ampliando el paquete *ade4* y lo vuelve capaz de analizar marcadores moleculares.

Otro paquete de R de amplio uso en los estudios de diversidad genética es *diveRsity*, que promete ser un paquete útil en la estimación y exploración de parámetros de genética de poblaciones y sus errores asociados (Keenan et al., 2013). En esencia, es un programa que calcula los mismos parámetros que *adegenet*; sin embargo, incorpora información para la validación estadística de sus resultados, de acuerdo con Keenan et al. (2013), calcula intervalos de confianza *Bootstrap* por locus y a nivel global.

Además de R, existen otros paquetes útiles en estudios de diversidad como el programa CERVUS, útil en el cálculo de frecuencias génica,  $H_o$ ,  $H_e$ , equilibrio Hardy-Weinberg y el coeficiente de consanguinidad. POPGEN ofrece cálculos similares, además de calcular el número observado y efectivo de alelos, y el flujo de genes. Los programas MEGA y STRUCTURE son útiles en la visualización gráfica, producen dendogramas y generan grupos de animales de acuerdo con su perfil genotípico usando estadística multivariada (Agung et al., 2019).

Otro programa usado es PLINK, líder en estudios de genética, posee desde análisis especializados como GWAS hasta el cálculo de parámetros básicos como lo hace *adeget*. PLINK es capaz de realizar cualquier método multivariado para la visualización de los datos, PCA, dendogramas, árboles filogenéticos, escalamientos multidimensionales y aprendizaje automático (Freitas et al., 2021).

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## CAPÍTULO 3. ALLELIC AND GENOTYPIC FREQUENCIES FOR LOCI ASSOCIATED WITH MEAT QUALITY IN MEXICAN BRAUNVIEH CATTLE

Tropical Animal Health and Production (2021) 53:307  
https://doi.org/10.1007/s11250-021-02757-5

REGULAR ARTICLES



### Allelic and genotypic frequencies for loci associated with meat quality in Mexican Braunvieh cattle

Mitzilin Zuleica Trujano-Chavez<sup>1</sup> · Jonathan E. Valerio-Hernández<sup>1</sup> · Rufino López-Ordaz<sup>1</sup> · Paulino Pérez-Rodríguez<sup>2</sup> · Agustín Ruiz-Flores<sup>1</sup>

Received: 23 December 2020 / Accepted: 2 May 2021  
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#### Abstract

The objective was to estimate allelic and genotypic frequencies for loci associated with meat quality in a Mexican population of Braunvieh cattle. Information was obtained from 300 animals genotyped with the Genomic Profile Bovine LD chip of 30K and 50K SNPs. After the final edition, including quality control, the data contained information for 12 loci of the CAPN1, CAPN3, CAPN5, CAPN14, DGAT1, DGAT2, TG, ANK1, and MADH3 genes. Allelic and genotypic frequencies and Hardy-Weinberg equilibrium were estimated with the Cervus 3.0.7 software. The studied population markers were in Hardy-Weinberg equilibrium, except for those associated with CAPN1, DGAT1, and MADH3. Frequencies higher than those reported for other breeds were found for genotypes associated with meat softness, higher marbling score, lower quantity of saturated fatty acids, and lower shear force (CAPN1 and DGAT2). There were similarities with frequencies reported for *Bos taurus* breeds for the CAPN3 and TG genes. For the DGAT1 and ANK1 genes, the frequencies of the desired genotypes were low. A marker for DGAT1 and another for MADH3 were monomorphic. The results of this study are encouraging in terms of the potential of the Braunvieh population studied for breeding programs aiming to increase meat quality. The breed has strengths that could be used either by crossbreeding to generate heterozygous animals or by selection to increase frequencies of valuable alleles.

**Keywords** Meat quality · DGAT1 · TG · CAPN1 · Genomic characterization · Hardy-Weinberg equilibrium

#### Introduction

Good quality beef is highly demanded. Meat quality is determined partially by visual, sensorial, and nutritional characteristics along with food safety and animal welfare that make it attractive for the consumer (Oliveira et al. 2014). Some of these characteristics are high marbling score, softness, juiciness, and tenderness (Ruiz et al. 2003).

Marker-assisted genomic selection (GWAS) could be a suitable option for achieving genetic progress in meat quality. This trait, as well as others of economic interest, according to Grisart et al. (2002), is of polygenic type and is under the control of several loci for quantitative traits (QTL). A long

generational interval limits traditional genetic improvement in beef cattle. Moreover, meat quality traits are usually measured in slaughtered animals, which delay the genetic progress and make the selection of young animals difficult.

If a gene is significantly associated with a trait or within a region harboring a QTL, it is called candidate gene (Long 2001). Significant effects for the CAPN1 gene for meat softness and marbling were reported in Brangus (Mazzucco et al. 2010), Canchim cattle (Curi et al. 2010), and Nelore (Tizioto et al. 2014). Additionally, Oliveira et al. (2014) in Nelore cattle found a close association of ANK1 with meat color and DGAT1 with increased marbling. The close association of marbling with DGAT1 was also reported in Nelore and Angeln cattle (Grisart et al. 2002; Sanders et al. 2006; Souza et al. 2010). In another study, Rubtsov and Lopina (2000) documented an important effect of ANK1 polymorphism on meat softness.

The frequency of polymorphisms associated with meat quality has been characterized genomically (allelic and genotypic frequencies for candidate genes) in Nelore populations in Brazil (Oliveira et al. 2014), Brangus in Argentina

✉ Agustín Ruiz-Flores  
aruizf@chapingo.mx

<sup>1</sup> Universidad Autónoma Chapingo, 56227 Chapingo, State of Mexico, Mexico

<sup>2</sup> Colegio de Postgraduados, 56264 Montecillos, State of Mexico, Mexico



(Mazzucco et al. 2010), and Braunvieh in Brazil (Curi et al. 2010). However, to our knowledge, this kind of study has not been conducted with Braunvieh cattle in Mexico. The objective of this study was to estimate allelic and genotypic frequencies for loci of candidate genes associated with meat quality in a Mexican Braunvieh population.

## Materials and methods

### Genotypic data

Hair samples were collected from 300 Braunvieh animals (236 female, 64 male) born from 2001 to 2016 on purebred farms belonging to the Mexican Association of Braunvieh Breeders from five herds in Eastern, Central, and Western Mexico. The samples were genotyped at GeneSeek (Lincoln, NE, USA, <http://genomics.neogene.com>). The chip used was Genomic Profile Bovine LD with 30,000 and 50,000 SNP markers for 150 different animals each.

### Loci identification

Markers or loci were identified using the genomic information available for *Bos taurus* at Gene (2020). The loci-gene allocation criterion was that the marker position was within the gene's allocation position range for the same chromosome. Only loci available in the genotyped samples were included.

Twelve loci of nine candidate genes reported in the literature to be associated with meat quality were used; these genes are shown in Table 1. Information of 30K and 50K genotypes was unified for CAPN1, CAPN5, CAPN14, DGAT2, TG, ANK1, and MADH3, each with one assigned marker, DGAT1 with two markers, and CAPN3 with three. The loci-gene allocation criterion was that the marker position was within the same chromosome's gene position range.

### Data analysis

For CAPN1 and DGAT1 analysis, information from 300 animals was used, while for the other seven genes, the analysis included 150 animals because the markers of the 30K marker chip did not contain the other loci in the study. Allelic and genotypic frequencies, as well as the Chi-squared test for Hardy-Weinberg equilibrium, were obtained with the Cervus 3.0.7 software package (Kalinowski et al. 2007).

The equation (Falconer and Mackay 1996) used by the software to test Hardy-Weinberg equilibrium is:

$$p^2 + 2pq + q^2 = p + q = 1$$

where  $p^2$  is AA homozygous genotypic frequency,  $2pq$  is heterozygote genotypic frequency (Aa), and  $q^2$  is aa

homozygous genotypic frequency. Also,  $p$  and  $q$  represent the A and a allelic frequencies.

## Results and discussion

The results of the characterization for meat quality candidate genes in the Braunvieh population studied are shown in Table 2. The allelic (alleles A and a) and genotypic (AA, Aa, and aa), general information of each marker, and its corresponding Chi-squared test to determine whether the population was or not in Hardy-Weinberg equilibrium (HW) are presented. The level of significance used in the test was 0.05; thus, values above 0.05 indicate that the population is in Hardy-Weinberg equilibrium.

Eight of the markers were in HW equilibrium (Table 2). However, markers associated with genes of major importance, such as CAPN1 and DGAT1, were not in HW equilibrium, and it could be due to either selection of the animals or sample size.

The studied population had a larger frequency of the desired genotype in the CAPN1 gene (AA frequency = 0.25) than other beef cattle breeds. In the literature it has been documented that genotypic variants (AA, Aa, and aa) did not significantly affect meat color. However, it has been reported that genotype AA is associated with higher marbling scores than the other genotypes (Li et al. 2013). These authors found frequencies of 0.00 to 0.11 for this genotype in *B. taurus* breeds (Angus had the highest frequency). The frequency for this genotype in Braunvieh is 0.25, Table 2, higher than in any previously mentioned breeds. According to Bonilla et al. (2010), in *Bos indicus* × *Bos taurus* cattle, the situation is similar; the frequency of genotype AA is on average 0.15. Similarly, genotype AA increases meat softness (Mazzucco et al. 2010). Therefore, allele A frequency in cattle populations should be as high as possible. In the present study, the results (0.5) were similar to those obtained (0.49) in Brangus (Mazzucco et al. 2010). In Brazilian Nelore and Braunvieh (Curi et al. 2010), genotype AA frequencies were essentially zero (0.000081 and 0.04489), results different from those of our study. These authors hypothesized that their values were due to the relatively small sample size used, 114 Nelore and 15 Braunvieh.

The Braunvieh population studied had similarities with other *B. taurus* breeds for the desirable genotypic frequencies of the gene CAPN3 (frequency range between 0.27 and 0.71). According to Barendse et al. (2008), the valuable genotype of the CAPN3 gene for greater softness is AA. These authors characterized the Red Belmont and Santa Gertrudis (frequencies for genotype AA, 0.44 and 0.32). Our study's frequency for this genotype was 0.27, close to that reported for Red Belmont and Santa Gertrudis. However, Brahman has a lower

**Table 1** Frequencies of homozygous and heterozygous genotypes reported in studies on beef cattle populations for genes associated with meat quality

| Gene                                                   | Symbol | Chrom | Genetic Group         | F <sub>HH</sub> | F <sub>HO</sub> | Reference                 |
|--------------------------------------------------------|--------|-------|-----------------------|-----------------|-----------------|---------------------------|
| Calpain 1 <sup>1, 2, 3, 5, 14</sup>                    | CAPN1  | 29    | Brangus               | 0.45            | 0.55            | Mazzucco et al. 2010      |
|                                                        |        |       | B. taurus×B. indicus  | 0.40            | 0.60            | Bonilla et al. 2010       |
|                                                        |        |       | Angus                 | 0.49            | 0.51            | Li et al. 2013            |
|                                                        |        |       | Charolais             | 0.21            | 0.79            |                           |
|                                                        |        |       | Hereford              | 0.06            | 0.94            |                           |
|                                                        |        |       | Simmental             | 0.33            | 0.67            |                           |
|                                                        |        |       | Nelore                | 0.02            | 0.98            | Curi et al. 2010          |
|                                                        |        |       | Braunvieh             | 0.13            | 0.87            |                           |
|                                                        |        |       | Canchim               | 0.17            | 0.83            |                           |
|                                                        |        |       | Angus×Hereford        | 0.41            | 0.59            | Corva et al. 2007         |
| Calpain 3 <sup>2</sup>                                 | CAPN3  | 10    | Brahman               | 0.42            | 0.58            | Barendse et al. 2008      |
|                                                        |        |       | Red Belmont           | 0.21            | 0.79            |                           |
|                                                        |        |       | Santa Gertrudis       | 0.31            | 0.69            |                           |
| Calpain 5 <sup>4, 2</sup>                              | CAPN5  | 15    | Nelore                | N/R             | N/R             | Tizioto et al. 2013       |
|                                                        |        |       | Blonde d'Aquitaine    | N/R             | N/R             | De Oliveira et al. 2017   |
|                                                        |        |       | Charolais             | N/R             | N/R             | Ramayo-Caldas et al. 2016 |
|                                                        |        |       | Limousin              | N/R             | N/R             |                           |
| Calpain 14 <sup>4</sup>                                | CAPN14 | 11    | Wagyu×Limousin        | N/R             | N/R             | Jiang et al. 2009         |
| Thyroglobulin <sup>5, 7, 8</sup>                       | TG     | 14    | Simmental             | 0.48            | 0.52            | Gan et al. 2008           |
|                                                        |        |       | Angus                 | 0.24            | 0.76            |                           |
|                                                        |        |       | Hereford              | 0.29            | 0.71            |                           |
|                                                        |        |       | Charolais             | 0.07            | 0.93            |                           |
|                                                        |        |       | Limousin              | 0.42            | 0.58            |                           |
|                                                        |        |       | Jinnan                | 0.43            | 0.57            |                           |
|                                                        |        |       | Nelore                | 0.00            | 1.00            | Fortes et al. 2009        |
|                                                        |        |       | Rubia Gallega×Nelore  | 0.04            | 0.96            |                           |
|                                                        |        |       | Brangus               | 0.40            | 0.60            |                           |
|                                                        |        |       | Braunvieh             | 0.39            | 0.61            |                           |
|                                                        |        |       | Angus                 | 0.98            | 0.02            | Li et al. 2013            |
|                                                        |        |       | Charolais             | 1.00            | 0.00            |                           |
|                                                        |        |       | Limousin              | 1.00            | 0.00            |                           |
|                                                        |        |       | Simmental             | 1.00            | 0.00            |                           |
|                                                        |        |       | Luxi                  | 0.22            | 0.78            | Yuan et al. 2013          |
| Diacylglycerol o-acyltransferase 1 <sup>5, 3, 6</sup>  | DGAT1  | 14    | Qinchuan              | 0.24            | 0.76            |                           |
|                                                        |        |       | Belgian blue          | 0.07            | 0.93            | Pannier et al. 2010       |
|                                                        |        |       | Blonde d'Aquitaine    | 0.33            | 0.63            |                           |
|                                                        |        |       | Salers                | 0.00            | 1.00            |                           |
|                                                        |        |       | Friesian              | 0.23            | 0.77            |                           |
|                                                        |        |       | Angus                 | 0.00            | 1.00            | Li et al. 2009            |
|                                                        |        |       | Jersey                | N/R             | N/R             | Dunner et al. 2013        |
|                                                        |        |       | Holstein              | N/R             | N/R             |                           |
|                                                        |        |       | Simmental             | N/R             | N/R             |                           |
|                                                        |        |       | Avileña-Negra Ibérica | N/R             | N/R             |                           |
| Diacylglycerol o-acyltransferase 2 <sup>13, 5, 2</sup> | DGAT2  | 15    | Piedmontese           | N/R             | N/R             |                           |
|                                                        |        |       | Marchigiana           | N/R             | N/R             |                           |
|                                                        |        |       | Limousin              | N/R             | N/R             |                           |
|                                                        |        |       | Charolais             | N/R             | N/R             |                           |
|                                                        |        |       | Bos Taurus            | N/R             | N/R             | Dunner et al. 2013        |
|                                                        |        |       | Nelore                | 0.47            | 0.53            | Oliveira et al. 2014      |
|                                                        |        |       | Charolais             | 0.13            | 0.87            | Horodyska et al. 2015     |
|                                                        |        |       |                       |                 |                 |                           |
| Malate dehydrogenase 3 <sup>2</sup>                    | MADH3  | 10    |                       |                 |                 |                           |
| Ankyrin 1 <sup>1, 5, 2, 9, 10, 11, 12, 14</sup>        | ANK 1  | 27    |                       |                 |                 |                           |

**Table 1** (continued)

| Gene | Symbol | Chrom | Genetic Group | F <sub>Ht</sub> | F <sub>Hh</sub> | Reference         |
|------|--------|-------|---------------|-----------------|-----------------|-------------------|
|      |        |       | Angus         | 0.19            | 0.81            | Aslan et al. 2010 |
|      |        |       | Charolais     | 0.29            | 0.71            |                   |
|      |        |       | Limousin      | 0.36            | 0.64            |                   |

*Symbol*, Symbol of the gene; *Chrom*, Chromosome; *F<sub>Ht</sub>*, Heterozygous genotype frequency; *F<sub>Hh</sub>*, homozygous genotype frequency; *N/R* = Not reported. Numeric superscripts in the names of genes indicate the associated meat quality characteristics: <sup>1</sup> Meat color, <sup>2</sup> Tenderness, <sup>3</sup> Shear force, <sup>4</sup> Amount of intramuscular fat, <sup>5</sup> Marbling, <sup>6</sup> Fat color, <sup>7</sup> Loin-eye area, <sup>8</sup> Rib-eye area, <sup>9</sup> Meat texture, <sup>10</sup> Juiciness, <sup>11</sup> Sarcomere length, <sup>12</sup> Cooking losses, <sup>13</sup> Fatty acid content, and <sup>14</sup> Softness

frequency, 0.19, than Braunvieh for this genotype (Barendse et al. 2008).

In the literature consulted, there were not found reports for allelic and genotypic frequencies for CAPN5, CAPN14, and MADH3 genes in beef cattle. However, in literature, there is ample evidence of association with meat quality traits, Table 1. In the present study, variability was found (Table 2) for CAPN5 genotypes. Hopefully, future research would allow a better characterization of Braunvieh regarding its allelic and genotypic frequencies. CAPN14 in Wagyu×Limousin crossbred cattle was found associated with seam fat (Jiang et al. 2009), as in Nelore (Tizioto et al. 2013). In the present study, for Braunvieh was nearly homozygous (aa = 0.93), the genetic variation is small. This could be either positive or negative, depending on further studies that elucidate the best genotype for CAPN14. Dunner et al. (2013) found an association of MADH3 with postmortem myofibrillar fragmentation

(tenderness). Frequencies for this gene were in a situation like that of the CAPN14 gene (AA = 1). Findings of future research will determine the convenience of this homozygosity.

Genotypic frequencies for the DGAT1 gene reported in literature have been different among breeds. Most of them had high proportions of the favorable genotype for marbling. According to our results, it was not present in Braunvieh. Li et al. (2013) reported an association of the DGAT1 gene with marbling in four European breeds. The heterozygous Aa genotype was associated with the best phenotype. In the same study, these authors reported for all four breeds, heterozygous frequencies above 0.98; therefore, *B. taurus* breeds would have the best DGAT1 genotype for marbling. This is contrary to our study's estimated frequencies, which coincide with those reported by Yuan et al. (2013) in Chinese commercial cattle with Aa frequencies below 0.24. Pannier et al. (2010) in Belgian Blue and Fortes et al. (2009) in Nelore, reported low

**Table 2** Allelic (A and a) and genotypic (AA, Aa, and aa) frequencies for loci associated with meat quality characteristics, and Hardy-Weinberg Equilibrium (HW) in the Braunvieh population studied

| SG     | Chrom | SNP Marker             | Position | Allele |      | Genotype |       |      | HW |
|--------|-------|------------------------|----------|--------|------|----------|-------|------|----|
|        |       |                        |          | A      | a    | AA       | Aa    | aa   |    |
| CAPN1  | 29    | UA-IFASA-1370          | 44085769 | 0.50*  | 0.50 | 0.25*    | 0.50  | 0.25 | b  |
| CAPN3  | 10    | BovineHD1000011720     | 37860216 | 0.71*  | 0.29 | 0.51*    | 0.41  | 0.08 | a  |
|        |       | BovineHD1000011723     | 37870339 | 0.47*  | 0.53 | 0.22*    | 0.50  | 0.28 | a  |
|        |       | Hapmap47063-BTA-62293  | 37852123 | 0.29*  | 0.71 | 0.09*    | 0.41  | 0.50 | a  |
| CAPN5  | 15    | BovineHD1500016485     | 57266887 | 0.40   | 0.60 | 0.16     | 0.48  | 0.36 | a  |
| CAPN14 | 11    | ARS-BFGL-NGS-10829     | 68687376 | 0.03   | 0.97 | 0.00     | 0.07  | 0.93 | a  |
| DGAT1  | 14    | ARS-BFGL-NGS-4939      | 1801116  | 0.94   | 0.06 | 0.88     | 0.11* | 0.00 | b  |
|        |       | Hapmap31987-BTC-062044 | 8425401  | 1.00   | 0.00 | 1.00     | 0.00* | 0.00 | b  |
| DGAT2  | 15    | UA-IFASA-5331          | 55185032 | 0.64   | 0.36 | 0.41*    | 0.46* | 0.13 | a  |
| TG     | 14    | BovineHD1400002314     | 8406694  | 0.64   | 0.36 | 0.41     | 0.46* | 0.13 | a  |
| ANK1   | 27    | ARS-BFGL-NGS-88722     | 36342691 | 0.23*  | 0.77 | 0.05*    | 0.36  | 0.59 | a  |
| MADH3  | 10    | BTB-00410322           | 13990943 | 1.00   | 0.00 | 1.00     | 0.00  | 0.00 | b  |

*SG*, Symbol of the gene; *Chrom*, Chromosome. The frequencies with the superscript (\*) indicates the desired alleles and/or genotypes by gene (improvement in meat quality). In the HW column, superscript <sup>a</sup> indicates there were non-significant differences ( $P > 0.05$ ) between the expected and the observed values in the Chi-squared test; this means the population is in Hardy-Weinberg equilibrium for the corresponding marker. On the contrary, <sup>b</sup> indicates the population is not in HW equilibrium



frequencies for the heterozygous Aa genotype, results like ours (0.07 in Belgian Blue and Nelore vs 0.00–0.11 in Braunvieh). Genetic improvement in Belgian Blue oriented to lean meat production could limit genotypes that code for higher production and deposit of fat, possibly explaining these DGAT1 frequencies in this breed.

Though less studied, the DGAT2 gene has significant effects on quality characteristics related to fat in *B. taurus* breeds. The Braunvieh population studied has the genetic potential for less seam fat. According to Li et al. (2009), the desired genotypes, in this case, were AA and Aa, associated with a low amount of seam fat. These authors studied three native Chinese breeds (Luxi, Jinnan, and Qinchuan), three crossbred (Charolais×Fuzhou, Limousin×Luxi, and Simmental×Jinan) and one *B. taurus* pure breed population (Angus). They found that genotype aa had frequencies above 0.98, results contrary to the 0.13 of our study, which means the total frequency of desirable genotypes is 0.87.

Braunvieh cattle exhibited frequencies for the desired TG genotype, Aa, like other *B. taurus* breeds. The TG gene is one of the most studied in beef cattle. According to Oliveira et al. (2014), in a study carried out in Brazil, the Braunvieh population had frequencies for Aa like those obtained in our study (0.39 vs 0.46). Gan et al. (2008) studied eight breeds; they reported that the desired genotype was the heterozygous, Aa, including the effects of six traits: live weight, carcass weight, carcass yield, seam fat, marbling, and loin- and rib-eye area. In the same study, the breed characterization showed that Simmental, Jinnan, and Limousin had the best frequencies (above 0.42), while Charolais had only 0.07 for the heterozygous genotypes. Our results in Braunvieh were like those reported for the first three breeds but higher than for Jinnan and Limousin (0.46 vs 0.43 and 0.42). On the other hand, Zhang et al. (2015), who studied the same breeds as Gan et al. (2008), found different results for TG, with high frequencies for homozygous genotypes, possibly due to the small number of animals used for estimations, on average 34 of each breed.

The favorable genotype for ANK1 (AA) is rare in Braunvieh compared with other breeds. The ANK1 gene, like TG, is one of the most studied and, according to Oliveira et al. (2014), the most valuable allele, considering intramuscular fat, rib-eye area, and cooking loss, is A. This means the best genotype is AA. The same authors reported a frequency of 0.2 for AA in Nelore, similar to findings in Charolais with a frequency of 0.32 (Horodyska et al. 2015). However, the differences in frequency for this gene in other breeds with Braunvieh are notorious, 0.2 vs 0.05.

In general, the Braunvieh population studied had higher frequencies of the desired genotypes than those of other breeds for CAPN1 and DGAT2 genes, mainly associated with fat content in meat. It also presents

similarities to CAPN3 and TG gene frequencies for most of *B. taurus* breeds, while for the DGAT1 and ANK1 genes, the frequencies of the most valuable genotypes were lower than those of other breeds. For the genes for which information was not found in literature, comparisons could not be made: CAPN5 presented a variation in the genotypes. In contrast, CAPN14 and MDH3 were monomorphic and homozygous; their interpretation is pending future research for valid conclusions.

Genetic frequencies indicate the genetic potential of a breed for economically important traits associated with meat quality. Only two of the 12 markers were monomorphic, indicating genetic variation in the studied population. The frequencies were high for valuable genotypes of genes associated with meat quality traits such as greater softness, marbling, lower amount of saturated fatty acids and shear force (CAPN1 and DGAT2). TG had similarities with other breeds, with high frequencies. This means that the population studied is potentially superior in the loin- and rib-eye area. The results were not favorable for genes associated with the yellow color of fat and less marbling; the desired genotype had frequencies below 0.07 (DGAT1 and ANK1).

The results in this study for meat quality traits, widely recognized as marbling and softness, were encouraging for the potential use of Braunvieh in beef cattle breeding programs. Selection could be used to increase alleles' frequency with high individual value to generate the homozygous genotypes favorable for meat quality traits.

**Supplementary Information** The online version contains supplementary material available at <https://doi.org/10.1007/s11250-021-02757-5>.

**Acknowledgements** The authors thank the Asociación Mexicana de Criadores de Ganado Suizo de Registro for allowing the use of their databases and the cooperation of the breeders for this study. Thanks also are extended to the Consejo Nacional de Ciencia y Tecnología, Mexico, for the financial support of the first author during her Master of Science studies.

**Author contribution** MZTC wrote the first version of the manuscript, JEVH and PPR assisted MZTC during the analysis. RLO and ARF conceived the project, all the authors contributed to obtain the final version, ARF served as the correspondence author.

**Funding** This work was supported by a grant from the Universidad Autónoma Chapingo, Mexico (DGIP-166701012).

## Declarations

**Conflict of interest** We declare no conflict of interest with any financial organization regarding the material discussed in the manuscript.

**Statement of animal rights** All applicable international, national, and/or institutional guidelines for the care and use of animals were followed. The manuscript does not contain clinical studies or patient data.



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**Publisher's note** Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

This chapter was sent to *Reproduction in Domestic Animals*

## CAPÍTULO 4. GENETIC DIVERSITY FOR REPRODUCTIVE TRAITS IN BRAUNVIEH CATTLE DETERMINED WITH SNP MARKERS

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### Reproduction in Domestic Animals

Genetic diversity in reproductive traits of Braunvieh cattle  
determined with SNP markers

|                                |                                                                                                                                                                                                                           |
|--------------------------------|---------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------|
| Journal:                       | <i>Reproduction in Domestic Animals</i>                                                                                                                                                                                   |
| Manuscript ID                  | RDA-OR-Oct-2021-0471                                                                                                                                                                                                      |
| Wiley - Manuscript type:       | Original Article                                                                                                                                                                                                          |
| Date Submitted by the Author:  | 29-Oct-2021                                                                                                                                                                                                               |
| Complete List of Authors:      | Trujano-Chavez, Mitzilin Zuleica; Universidad Autonoma Chapingo Ruiz-Flores, Agustin; Universidad Autonoma Chapingo López-Ordaz, Rufino; Universidad Autonoma Chapingo Pérez-Rodríguez, Paulino; Colegio de Postgraduados |
| Subject Area:                  | Animal breeding < General reproduction, Genetics < General reproduction, cattle < Species:                                                                                                                                |
| Editor Preference Recommended: | Prof. Dr. E. Martinez-Garcia, Murcia, Spain                                                                                                                                                                               |
| Editor Preference Opposed:     | Prof. Dr. F. Almeida, Belo Horizonte, Brazil                                                                                                                                                                              |
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**1 Genetic diversity in reproductive traits of Braunvieh cattle determined with SNP**  
**2 markers**

3 Mitzilin Zuleica Trujano-Chavez<sup>a</sup>, Agustín Ruiz-Flores<sup>a</sup>, Rufino López-Ordaz<sup>a</sup>, Paulino  
4 Pérez-Rodríguez<sup>b</sup>

5 <sup>a</sup> *Posgrado en Producción Animal, Universidad Autónoma Chapingo, Carretera Federal*  
6 *México-Texcoco Km 38.5, 56227, Texcoco, Estado de México, México.*

7 <sup>b</sup> *Socio Economía Estadística e Informática, Colegio de Postgraduados, Carretera*  
8 *Federal México-Texcoco Km 36.5, 56230, Texcoco, Estado de México.*

9  
10 Corresponding author: Paulino Pérez-Rodríguez, Email: perpdgo@gmail.com

11  
12 Corresponding author ORCID: 0000-0002-3202-1784

**14 Abstract**

15 Braunvieh is an important dual-purpose breed in the Mexican tropics. The study of its  
16 genetic diversity is key to implementing genetic improvement programs. This study was  
17 conducted to determine genetic diversity of reproductive traits in a Mexican Braunvieh  
18 beef cattle population using single nucleotide polymorphisms in candidate genes.

19 Information from 24 genes with 52 intra-genic loci reported in literature to be associated  
20 with productive life, pregnancy rate, and cow and heifer conception rate of 150 animals  
21 was considered. Observed heterozygosity (Ho) revealed high genetic diversity for the  
22 studied traits, Ho=0.42, relative to that of other populations of the same breed. Cluster  
23 analyses were carried out using the Ward and K-means algorithms. These analyses



revealed genetic diversity that formed at least two groups of animals by genotype. These results coincide with what was observed in the biplot of non-metric multidimensional scaling. Allelic frequencies for the groups formed with the K-means algorithm were estimated as well. It was found that this grouping strategy allowed visualization of distant groups by genotype but not by favorable alleles in all the loci. In addition, the genes *CSNK1E*, *DNAH11*, *DSC2*, *IBSP*, and *OCN* affected most of the traits and they were highly informative. Therefore, they represent a potential resource for selection of the traits studied in Braunvieh. The analyses showed that the Mexican Braunvieh population has a high level of genetic diversity, arguably due to decades-long adaptation to the Mexican tropics.

**Key words:** Brown Swiss, candidate gene, cluster analysis, genetic variability.

## Introduction

One of the first objectives in animal breeding was to increase production efficiency per animal (Fleming et al., 2018). Production of animals of high genetic merit allowed improvement of economically important traits, such as milk production in dairy cattle or average daily gain in chickens. However, some traits of similar or higher importance were dismissed. According to Veronese et al. (2019), for years genetic selection in cattle ignored the improvement of reproductive traits.

Today, the strategy in most dairy and dual-purpose herds has emphasized selection of productive and reproductive traits jointly to make the production system more efficient. Since 2000, genetic and genomic selection has included, among others, characteristics such as cow and heifer conception, and positive production results have

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3 48 correlated highly with the animals' genetic merit (Veronese et al., 2019). Recently, single  
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5 49 nucleotide polymorphisms (SNP) have become a widely used resource for genomic  
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8 50 selection. One of the reasons is that the cost of genotyping has decreased to a cost-  
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10 51 effective level.

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12 52 Genetic diversity is a key element in any genetic improvement program. Breeds  
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14 53 adapted to harsh environments are usually more genetically diverse because they need  
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16 54 to be versatile to face changes when they are moved from one environment to another  
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18 55 (Xia et al., 2018). One of the most demanding production systems for cattle production  
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20 56 is the dual-purpose system in tropical regions. The breeds that have had satisfactory  
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22 57 results in these conditions are *Bos indicus*, such as Indubrasil (Peixoto et al., 2021) or  
23  
24 58 Guzerat (Campos et al., 2017). Recently Braunvieh has experienced an important boom  
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26 59 in these production systems because of its good results for beef production. Braunvieh  
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28 60 is a breed that is well adapted to cold climates (Bhati et al., 2020). It has been  
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30 61 successfully used in dual-purpose systems as the paternal breed (Rojo-Rubio, 2009).  
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32 62 Their successful adaptation to tropical conditions suggests that the Mexican population  
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34 63 of Braunvieh has important genetic diversity with potential use in genetic improvement  
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36 64 programs for reproductive traits in dual-purpose systems.

37  
38 65 Genetic diversity studies have been conducted in Braunvieh and its derivative  
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40 66 breeds, aiming to unravel the genetic basis of its adaptation to tropical environments.  
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42 67 Moscarelli et al. (2020) found important genetic differences among several Braunvieh-  
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44 68 derived breeds. Moreover, Bhati et al. (2020) found that Braunvieh has higher genetic  
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46 69 diversity than other dual-purpose *Bos taurus* breeds. The objective of our study was to  
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48 70 characterize genetically a Mexican Braunvieh population for productive life, pregnancy  
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50 71 rate, and cow and heifer conception rate, through an analysis of genetic diversity  
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3 72 focusing on genotypes and favorable alleles for intra-genic SNPs of candidate genes,  
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5 73 using expected and observed heterozygosity indexes, grouping algorithms, analysis of  
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8 74 non-metric multidimensional scaling, and estimation of allele frequencies.  
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10 75

## 11 12 76 **Material and methods**

### 13 14 77 ***Source of information***

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17 78 Genomic information was obtained from hair samples from 150 animals born  
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19 79 between 2001 and 2016 on five farms belonging to the Mexican Association of  
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21 80 Braunvieh Purebred Breeders in Eastern, Central and Western Mexico. The samples  
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23 81 were genotyped at GeneSeek (Lincoln, NE, USA, <http://genomics.neogene.com>). The  
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25 82 chip used was the Genomic Profile Bovine LD with 50,000 SNP markers.  
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### 29 30 84 ***Genotype quality control***

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33 85 For data edition before genotype analyses, quality control was performed. The  
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35 86 missing genotypes were imputed using the observed allele frequencies. The imputation  
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37 87 method consisted of assigning an allele, according to the probability of a possible type of  
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39 88 polymorphism for a certain marker in the whole population. Additionally, we considered  
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41 89 a 0.05 threshold for the Minor Allele Frequency to discard non-informative markers or  
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43 90 markers with errors in the genotyping.  
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### 47 48 92 ***Statistical analyses***

#### 49 50 93 ***Identification of associated and informative loci***

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52 94 The candidate genes associated with reproductive traits in *Bos taurus* were  
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54 95 identified. The position ranked by base pairs were obtained in the Gene Library (Gene,  
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2021). With this information, the intragenic loci or SNP markers were searched; the criterion for the loci-gene association was that they should be located within the position rank of the gene for the same chromosome. Only the loci available in the genotype information were included. The analyses considered 52 SNP markers of 24 candidate genes reported in the literature associated with pregnancy rate (PR), heifer (HCR) and cow conception rate (CCR), and productive life (PL).

All the analyses were performed using the R software version 4.0.4 (R Core Team, 2021). The 10 or 12 most informative markers associated, according to the literature consulted, with each of the studied traits were determined. The Shannon index (H; Tables 1 and 2) was used. This index allows identification of the most informative markers (Sherwin, 2010). The index is defined as:

$$H = - \sum_{i=0}^2 p_i \log p_i,$$

where  $i = 0, 1, 2$  refers to the genotype ( $AA = 0, AB = 1, BB = 2$ ), and  $p_i$  is the corresponding allelic frequency and  $\log(\cdot)$  denotes the natural logarithm function. This index was calculated for each of the loci. Empirical distribution was obtained, and the 10 or 12 most informative markers were selected. The highest values for H were associated with the highest diversity for the marker.

#### *Cluster analyses*

The groups formed by their genetic diversity were generated using hierarchical grouping based on the Euclidian distance matrix as a measure of dissimilarity and the Ward distance as a measure of proximity between groups (Ward, 1963). The analysis was performed using the cluster function included in the stats package of R (R Core



Team, 2021). Additionally, data were also clustered using the K-means algorithm, using the `kmeans` function included in the `stats` package in R. In both cases the objective was to generate groups of animals with similar genotypic profiles for each of the studied traits. The 10 most informative markers were used. K-means is a non-hierarchical method of re-assignment that allows grouping similar individuals, the method finds K non-overlapped groups (Wu, 2012). According to Mesquita et al. (2017), the Euclidean distance  $\eta_{ij}$  between two vectors  $x_i$  and  $x_j$ , of dimension D, is given by:

$$\eta_{ij} = z_{ij}^{1/2} = \sqrt{\sum_{d=1}^D (x_{i,d} - x_{j,d})^2},$$

where  $z_{ij} = \|x_i - x_j\|_2^2 = \sum_{d=1}^D (x_{i,d} - x_{j,d})^2$  is the squared distance between  $x_i$  and  $x_j$ , with  $i, j = 1, \dots, n$  and  $n$  is the number of individuals. In the context of our analysis, the vectors  $x_i$  and  $x_j$  represent the genotypic information recoded for additive effects for individuals  $i$  and  $j$ .

To validate the results found using hierarchical grouping and the K-means method, the data was also analyzed using the Partition Around Medoids (PAM) algorithm (Lengyel and Botta-Dukát, 2019). PAM yields a statistic and a graph which determines grouping quality on a global and individual level using silhouette width and average silhouette width for each group and average silhouette width including all individuals. We performed the analysis using the `pam` function included in the `cluster` package (Maechler et al., 2019) in R. Average values of silhouette width by individual and by group were obtained. The desirable values are close to 1, meaning that on average the objects are close to others in the same group and their classification is

140 correct. On the other hand, values close to -1 indicate that the classification is

141 undoubtedly incorrect (Lengyel and Botta-Dukát, 2019).

142

143 *Non-metric multidimensional scaling*

144 NMDS was used for graphic representation in the bi-dimensional space of the

145 distance matrices (per trait), using the *vegan* library in R (Oksanen et al., 2020). The

146 stress value proposed by Kruskal (Dexter et al., 2018) was obtained. This metric

147 estimator indicates how well the algorithm has managed to arrange the points in the

148 ordination while preserving the rank-order distance. The desirable stress values range

149 from 0 to 0.2. Stress values > 0.35 indicate that the samples are randomly placed in the

150 graphic representation and are also considered poor and potentially uninterpretable

151 (Dexter et al., 2018). This multi-variant analysis was combined with the K-means results

152 to visualize graphically the clusters and the animal with the representative genotype of

153 each group (the closest to the centroid).

154

155 *Heterozygosity*

156 Levels of expected ( $H_e$ ) and observed ( $H_o$ ) heterozygosity for the 52 initial

157 markers of the studied population were obtained using the software Cervus 3.0.7

158 (Kalinowski et al., 2007). Standard deviations were estimated through the non-

159 parametric Bootstrap method for 10,000 replicates using the *boot* function included in

160 the package *boot* in R (Canty and Ripley, 2021).

161

162 *Allelic frequencies and proportion test*

163 The allelic frequencies for each group formed with the K-means algorithm were  
 164 obtained for each studied trait using the software Cervus 3.0.7 (Kalinowski et al., 2007).  
 165 To determine whether significant differences existed between allelic frequencies for  
 166 desirable alleles of the groups formed, and those of representative animals, proportion  
 167  $X^2$  tests were carried out using the *chisq.test* included in the *stats* package in R (R Core  
 168 Team, 2021). The null hypothesis was that non-significant differences exist between  
 169 proportions of two or more groups (Shih and Fay, 2017). Graphs were generated to  
 170 visualize proportions for the frequencies of favorable alleles of the most informative  
 171 genes in the database that affect 3 to 4 of the studied traits: *CSNK1E*, *DNAH11*, *DSC2*,  
 172 *IBSP*, and *OCLN*.

## 174 **Results and Discussion**

### 175 ***Identification of associated and informative loci***

176 The 10 most informative loci associated with the studied traits are shown in  
 177 Tables 1 and 2. For each gene, the favorable allele reported in literature is in  
 178 parentheses. For some of the genes, there are over-dominance reports; therefore, the  
 179 favorable genotype is the heterozygote. However, genotypes are not transmitted from  
 180 one generation to the next. Therefore, even though the effect of allele interaction is  
 181 relevant in crossbreeding breeding systems, our study on genetic diversity focuses on  
 182 alleles whose additive effect is transmitted from one generation to another.

### 183 ***Heterozygosity***

184 Genetic diversity indexes ( $H_e$  and  $H_o$ , Table 3) are key parameters in genetic  
 185 improvement of populations. They were used in our study to determine the level of  
 186 genetic diversity in the population studied. The genetic diversity measured as heterosis



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3 187 was quite similar among the 21 markers. The frequency of heterozygote individuals in  
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5 188 the population was close to 0.5, which indicates that the genetic variation corresponds  
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8 189 with what was expected, according to Hardy-Weinberg Equilibrium (HWE). In fact, all the  
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10 190 markers in the study at  $p > 0.0003$  (Bonferroni correction) were in HWE.

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12 191 These results suggest that genetic diversity of reproductive traits in the Mexican  
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14 192 Braunvieh population is higher than the average of the breeds derived from the original  
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16 193 Braunvieh from Switzerland. Moscarelli et al. (2020) reported values close to 0.35 for  
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18 194 European populations of original Braunvieh, Braunvieh, Brown Swiss, and Italian Brown.  
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20 195 This suggests an important difference from our findings,  $H_o = 0.42$  (Table 3). Moscarelli  
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22 196 et al. (2020) found that animals of the same breeds that stayed in Switzerland have a  
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24 197 higher  $H_o$  than those adapted to other environments, such as the Italian Brown, possibly  
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26 198 because of conservation of purebred systems. In the population studied the opposite  
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28 199 occurred: heterozygosity was higher, possibly due to its adaptation to the tropical  
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30 200 environment, which is very different from that in which Braunvieh originated.

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32 201 The  $H_o$  in the Braunvieh population studied for reproductive traits coincides with  
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34 202 the findings for other breeds adapted to harsh environments. Campos et al. (2017)  
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36 203 reported  $H_o$  for nine *Bos indicus* breeds; their estimates ranged from 0.32 to 0.39,  
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38 204 values higher than those found for the breeds related to Braunvieh previously discussed.  
39  
40 205 These results coincide with the findings of our study, although the  $H_o$  of the Mexican  
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42 206 population is still higher. Xia et al. (2018) reported  $H_o$  estimates for more than 40 Asian  
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44 207 cattle breeds. These authors observed that a value close to 0.5 was associated with  
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46 208 breeds, such as Kazakh, Mongolian, and Anxi, adapted to the extreme environments of  
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48 209 Russia and Mongolia. These authors reported estimates of 0.25 for breeds adapted to  
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50 210 temperate climates. In another study, Peixoto et al. (2021) reported values for  $H_o$

211 between 0.47 and 0.85 in Brazilian Guzerat, which is adapted to tropical environments.  
212 According to these authors, high genetic variation is typical of Creole populations or  
213 those subject to HWE. This could be the reason that  $H_o$  for the Braunvieh population of  
214 our study is like that of breeds adapted to extreme climates. The Mexican Braunvieh  
215 population has had more than a century of adaptation to tropical conditions.

216

#### 217 ***Clustering using the Ward and K-means algorithms***

218 The average silhouette width for all the traits obtained with the Ward method was  
219 0.13, while it was 0.18 using the K-means method. These estimates suggest that  
220 clustering with the K-means algorithm is more adequate than with the Ward method.  
221 Although these estimates were low, they were positive in all the cases. Therefore, they  
222 can be considered acceptable values that reveal clear genotypic differences between  
223 the groups formed with the algorithms (Lengyel and Botta-Dukát, 2019).

224 Both methods of analysis yielded the same number of formed clusters, two for  
225 PR, HCR, and CCR, and three for PL. The circular dendrograms obtained from the  
226 dissimilarity matrix with the Ward algorithm for the studied traits, allowed grouping the  
227 150 individuals of the study population into different groups (Figure 1).

228 Clustering algorithms are a useful tool in genetic diversity studies. The findings of  
229 the present study, relative to the clustering power of the methods for characterizing  
230 genetic diversity, coincide with Campos et al. (2017), who used principal components  
231 analysis (PCA) in Zebu cattle in Brazil. They also agree with Öner et al. (2017), who  
232 used Unweighted Pair Group Method using Arithmetic Averages to create a  
233 phylogenetic tree between Holstein populations, and with Kim et al. (2018), who used  
234 the same procedure for Korean breeds.



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3 235 The results of clustering suggest that the diversity found is sufficient to create at  
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5 236 least two groups by trait, which differ in the proportion of animals and in the combination  
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8 237 of favorable alleles. In the Figure 1 dendrogram for PR, animal 141 (cluster I) is  
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10 238 classified in the smallest group, which contains animals with predominantly BB  
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12 239 genotype. In contrast, the largest group was made up of animals in which the AA  
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14 240 genotype predominates (Table 4). For PL, three clusters were formed; those containing  
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16 241 animals 123 (cluster II) and 133 (cluster III) have the same proportion of genotypes.  
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18 242 However, they differ in genotypes for the genes DSC2, HSD17B12, IBSP, LHCGR, and  
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20 243 OCLN. These two groups differ from cluster I (132 is the representative animal), which  
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22 244 does not possess BB genotypes, but only AA and AB genotypes in a 2:3 proportion  
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25 245 (Table 4 and Table 1). The dendrograms for CCR (d) and HCR (c) show that the  
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27 246 groupings are similar. For CCR, the largest group (cluster II), according to the  
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29 247 representative animal 112, contains a greater proportion of BB genotypes, while cluster I  
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31 248 (animal 135), contains a greater proportion of AB genotypes. For HCR, cluster I (animal  
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33 249 54) contains mostly AB genotypes, and AA genotypes are absent. On the other hand,  
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36 250 animal 93 is representative of cluster II, which includes mostly AB genotypes.  
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40 251 Table 4 shows the different groups formed with K-means. Given the large number  
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42 252 of animals studied, only the five closest to the centroid and the representative genotype  
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44 253 of the individual closest to the centroid are reported. In addition, the percentages of  
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46 254 desirable genotypes and alleles of each individual are shown to detect the group of  
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49 255 animals with mostly favorable genotypes that would be useful in selection.  
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51 256 There are few significant differences in the proportion of desirable alleles and  
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53 257 genotypes between the groups formed with the K-means algorithm due to a  
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56 258 compensatory effect in the combination of alleles (Moscarelli et al., 2020). In Table 4, it  
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could be observed that the proportion of desirable alleles does not differ significantly (Tables 1 and 2) between groups. An exception was for PL, where cluster III has the lowest significantly different proportion of favorable alleles.

The few differences between proportions are attributed to a compensatory effect, a consequence of analyzing the genes as a set to obtain the proportions per group. This procedure was due to the high degree of dissimilarity in genetic variation between markers and genes. Although the main reason for compensation could be that grouping with the K-means method places the animals into different groups because their genotypes are different, generating specific combinations; for example, for PR, cluster I includes a larger number of animals with BB genotypes than cluster II.

These differences do not necessarily mean that any of the groups has a higher number of desirable genotypes or alleles. Alleles vary as a function of the genes; for example, in PR for the gene *BCAS* the desirable allele is A; however, for the *CSNK1E* gene it is B. This means that cluster I contains the favorable allele for *CSNK1E*, but not for *BCAS*, and vice versa for cluster II. The same holds true for most of the cases. That is, the proportions of favorable alleles for all the groups are close to 0.5 (Table 4) due to the compensatory effect determined by the combination of alleles with which this grouping method works (Moscarelli et al., 2020), but it is not related to the number of favorable alleles.

The grouping methods suggest the existence of genetic diversity that would be useful in improvement programs. Despite the compensatory effect previously described, the genetic variation found is good, given that the proportions of favorable genes are important, even though they do not appear assembled together for all the genes. According to Peixoto et al. (2021), the high variation and the eventual formation of

groups is an indication of diversity conservation and assures breed sustainability over generations. Today, genetic conservation of adapted or local breeds is a relevant concern due to their contribution to sustainable selection of genotypes.

#### ***Non-metric multidimensional scaling***

In Figure 2, the bi-dimensional representations obtained with NMDS for the studied traits are shown. To visualize the grouping by genotypes, the K-means classification was included in the graphic, highlighting each of the groups found and the representative animals of each group (Table 4).

The Kruskal stress values for PR (0.22), CCR, and HCR (0.24) are acceptable given they are not greater than 0.35, while the value for PL (0.15) permits graphic visualization of what NMDS offers (Dexter et al., 2018). NMDS is a useful tool to visualize the groups of animals with different genotypes found by the K-means algorithm. Figure 2, from the NMDS analysis, shows that there were no individuals classified in groups different from those assigned by the K-means method. Similarly, the representative animals are close to the center of the graphic representation of their groups. Although on the borders of the groups some animals are not clearly classified, the groups are well delimited with respect to their neighboring groups.

NMDS is a tool widely used in genetic diversity studies. Like the findings of our study, satisfactory results have been found in the graphic representation of the method, for example, the study of Moscarelli et al. (2020) with breeds derived from the original Braunvieh and the study of Senczuk et al. (2020) where NMDS was used to visualize bi-dimensionally differences among cattle breeds from the Alpine arc.



### 307 ***Allele frequencies***

308 In Mexico, studies using massive genomic information are scarce. According to  
 309 Mrode et al. (2019) the main reason is the limited resources available to invest in the  
 310 genotyping of animals in developing countries. These authors point out that the genetic  
 311 improvement infrastructure of these countries cannot be compared with the advanced  
 312 infrastructures of developed countries, whose genomic databases are made up of  
 313 millions of animals. Given these limitations, the studies of Zepeda-Batista et al. (2019)  
 314 and Trujano-Chavez et al. (2021), used 300 animals to characterize a Mexican  
 315 Braunvieh population, the same used in our study, but with a more limited number of  
 316 animals since the loci found for reproductive traits were only available for 150 animals.

317 The sample size used in our study, checked out using the *pwr.p.test* function from  
 318 the *pwr* package (Champely, 2020), was sufficient to detect differences between groups.  
 319 The sample size (150) of our study, according to Canals and Canals (2019), was  
 320 adequate to find significant differences with a 0.05 significance level obtaining a power  
 321 of the proportion test of 0.92. Other studies like ours found relevant results with even  
 322 fewer animals. The studies of Agung et al. (2019), Terentjeva et al. (2021), and Xia et al.  
 323 (2018) had average sample size of 22, 20, and 25 animals, respectively.

324 Table 5 shows the proportion tests to determine the significant differences  
 325 between allelic frequencies of desirable alleles per group and gene. Since the  
 326 genotypes are non-heritable, only allelic frequencies were considered. The genes  
 327 *DZIP3*, *LHCGR*, *IBSP*, *APBB1*, *GOLGA4*, and *DSC2* were reported with dominance and  
 328 over-dominance effects in *Bos taurus* (Cochran et al., 2013; Ortega et al., 2015).  
 329 Therefore, the allele considered desirable was that with the greatest additive value for

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3 330 the different traits. Furthermore, the frequencies per gene were considered because the  
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5 331 frequencies of the loci or SNP markers within each gene were not significantly different.  
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8 332         Given that the general results (Table 4) for favorable alleles at the genotype level  
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10 333 (10 and 12 loci) were non-conclusive because of the compensatory effect, the effect of  
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12 334 clustering on changes in allelic frequencies was analyzed at the gene/marker level. At  
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14 335 the allelic frequency level per gene, there were significant differences ( $p < 0.05$ ) between  
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16 336 groups as a product of clustering for PR; group II had the largest number of alleles with  
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18 337 favorable effect. The group frequencies for *DSC2*, *LHGR*, and *OCN* were significantly  
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20 338 different. That is, the classification of K-means was a function of these three genes.  
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23 339 Group II had the largest number of favorable alleles for *DSC2* and *LHCGR*, while group I  
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25 340 had the highest number for *OCN*.  
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28 341         Group I had the lowest frequency of favorable alleles for PL and group II the  
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30 342 highest. As was previously seen in the first section of grouping, group I for PL had the  
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32 343 lowest proportion of favorable alleles when the analysis was performed at the allele  
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34 344 combination level (Table 3) and at the gene level (Table 5). In group II alleles of the  
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36 345 genes *FSHR*, *HSD17B12*, *IBSP*, and *LHCGR* had higher frequencies. Although group III  
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38 346 was better than group I, it had frequencies with significant differences for only *DSC2* and  
39  
40 347 *OCN*. Therefore, group II would be selected when selection is intended for PL.  
41  
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43 348         The differences between allelic frequencies of the CCR groups are not enough to  
44  
45 349 determine that either of the groups is better. In contrast with the traits previously  
46  
47 350 discussed, the results for CCR are not conclusive by themselves. This is because in  
48  
49 351 both groups two genes exist with significantly higher frequencies (Table 5), and the  
50  
51 352 average for the frequencies is not different ( $p > 0.05$ ) because of the compensatory effect.  
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54 353 Therefore, for this trait in particular, the animals should be selected based on the other  
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traits, that is, the animals in favorable groups for the other high-frequency traits for the genes they have in common. For example, if group I were selected for PL, group I animals will be selected indirectly for CCR since *DSC2* is a common gene. For HCR, the results were different from those for CCR. Group I is the selection candidate to improve this trait. Unlike its homologous trait, in HCR group I possesses significant differences for two genes (*DNAH11* and *DSC2*), while group II has significant differences only for gene *GOLGA4*.

Some of the genes studied have more relevance for joint selection of animals. Genes *CSNK1E*, *DNAH11*, *DSC2*, *IBSP*, and *OCN* affect two to three of the studied traits. Therefore, selecting exclusively for these genes could have a generalized impact where selection is for multiple traits. Gene *DSC2*, which affects the four studied traits, could be a direct selection objective to improve reproductive traits. The results of the proportion test for the groups formed by each of the traits for these genes of the higher importance is shown in Figure 3.

Although for the most important genes some of the frequencies of favorable alleles are not significantly different between groups by trait, the results suggest that it is possible to leverage some of the groups of animals by their frequency of favorable alleles. This can be observed for example in Figure 3: for *DSC2* group II for PR, group I for CCR and HCR, and group III for PL. Another gene with differences in frequencies of favorable alleles per group is *OCN*. Group I for PR could be useful, as is group II for CCR and III for PL.

The genetic diversity found indicates that even though not all the animals have favorable alleles, many of them have very desirable alleles. This information could be enough to start genetic improvement programs for reproductive traits in Braunvieh. The



genes considered in this study are the product of selection for variation of, not for addition to, the genetic merit of the animals. In the literature reviewed nearly 24 genes associated with the traits studied were found. However, the Shannon index allowed selection of only the most informative markers of those with greater variation. For this reason, the results of our study may be different from those reported in research with genes of major effects, such as the study of Ortega et al. (2015), where the genes *ACAT2*, *AP3B1*, and *OCN* had a greater impact on the genetic value of Holstein animals for reproductive traits than *DSC2*. Gene *DSC2* is considered the most important of our study because its variation allowed classifying the animals in different groups, and it affects the four studied traits, and not because it had the greatest effect as compared with the other genes.

389

#### 390 ***Final considerations***

The results of our study suggest the presence of genetic diversity for the studied traits in the population analyzed. Genetic diversity studies have shown that there are different Braunvieh lines adapted to different environments. This process has had an important effect on the population variation (Moscarelli et al. 2020; Senczuk et al., 2020). In Mexico, the breed has been slowly introduced into dry and humid tropical areas aiming for its use as a dual-purpose breed. This has modified its allelic combinations, as well the  $H_o$ , relative to other Braunvieh populations of the world. Braunvieh is widely in commercial dairy operations because of its milk and beef quality conferred by its Swiss cattle origins, the world's most ancient (Rojo-Rubio et al., 2009). The  $H_o$  estimates found in this study suggest adaptation of the Braunvieh population to the Mexican tropics since these values differ from those of its direct ancestors and

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3 402 coincide with those obtained in breeds adapted to extreme conditions. Additionally, the  
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5 403 groups formed by the K-means algorithm and the differences among the allele  
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8 404 combinations support the existence of genetic variation for reproductive traits, allowing  
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10 405 the possibility to select animals for specific reproduction objectives to improve PR, PL,  
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12 406 CCR or HCR, or through the gene *DSC2* or any of the representative genes, using  
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14 407 group allelic frequencies.

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17 408 The results of this study constitute a first approach to genetic characterization for  
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19 409 reproductive traits of cattle in Mexico. They are satisfactory because the presence of  
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21 410 genetic diversity is considered an opportunity to implement selection and/or  
22  
23 411 crossbreeding plans to improve reproductive performance. Likewise, genetic diversity is  
24  
25 412 essential for conservation of populations of genotypes adapted to different  
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27 413 environments. Studies on diversity can provide the information necessary to identify  
28  
29 414 genes associated with traits of interest, in this case, for animals well-adapted to  
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31 415 conditions different from those of their origin. Genetic diversity is not a concept  
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33 416 associated only with a great number of loci, as in this study where only 52 SNP markers  
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35 417 were used. Other studies have been carried out using information of only one gene with  
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37 418 few alleles. For example, Takeshima et al. (2018) carried out a study on genetic  
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39 419 diversity in *Bos indicus* cattle in South America using information of only the gene *BoLA-*  
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42  
43 420 *DRB3*.

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#### 47 48 49 422 **Acknowledgements**

50  
51 423 Authors thank the Consejo Nacional de Ciencia y Tecnología, México, for the financial  
52  
53 424 support to the first author during her Master of Science studies. The authors also thank  
54  
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3 425 the Asociación Mexicana de Criadores de Ganado Suizo de Registro for allowing the  
4  
5 426 use of their databases and the breeders for their kind cooperation in this study.  
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7 427  
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10 428 **Ethics approval**  
11  
12 429 All applicable international, national, and/or institutional guidelines for the welfare and  
13  
14 430 use of animals were met.  
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17 431  
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19 432 **Author contributions**  
20  
21 433 MZTC drafted the first version of the manuscript, PPR supported MZTC during the  
22  
23 434 analysis, ARF and RLO conceived the project. All the authors contributed to obtain the  
24  
25 435 final version, PPR acted as the author for correspondence.  
26  
27 436  
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29  
30 437 **Conflict of interest**  
31  
32 438 None of the authors has any conflict of interest to declare.  
33  
34 439  
35  
36  
37 440 **Financial support**  
38  
39 441 This work was in part supported by a grant from the Universidad Autónoma Chapingo,  
40  
41 442 México (DGIP-166701012).  
42  
43 443  
44  
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46 444 **Data availability**  
47  
48 445 Upon reasonable request to the author for correspondence.  
49  
50 446  
51  
52  
53 447 **Author ORCIDs**  
54  
55 448 Mitzilín Zeleica Trujano-Chavez 0000-0002-3242-1040  
56  
57  
58  
59  
60



449 Rufino López-Ordaz 0000-0002-1825-0097

450 Agustín Ruíz-Flores 0000-0001-8267-2107

451 Paulino Pérez-Rodríguez 0000-0002-3202-1784

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**Table 1.** Top 12 and 10 single nucleotide polymorphisms (SNP) associated with pregnancy rate and cow productive life in Braunvieh Cattle.

| Pregnancy rate         |                        | Productive life        |                       |
|------------------------|------------------------|------------------------|-----------------------|
| Gene (DA) <sup>1</sup> | SNP                    | Gene (DA) <sup>1</sup> | SNP                   |
| APBB1 (A)              | BovineHD1500013531     | AP3B1 (A)              | BovineHD1000003029    |
| BCAS1 (A)              | BovineHD1300023602     | DSC2 (B)               | BovineHD2400007184    |
| BOLA-DMB (B)           | Hapmap60475-rs29022896 |                        | BovineHD2400007183    |
| CSNK1E (B)             | Hapmap39945-BTA-75021  | FSHR (B)               | ARS-BFGL-NGS-5623     |
| DNAH11 (A)             | BovineHD0400008600     |                        | Hapmap45323-BTA-90907 |
|                        | Hapmap38265-BTA-96973  | HSD17B12 (A)           | ARS-BFGL-NGS-4967     |
| DSC2° (A)              | BovineHD2400007184     |                        | BovineHD1500021362    |
|                        | Hapmap52659-ss46526625 | IBSP° (A)              | BovineHD0600010277    |
|                        | BovineHD2400007183     | LHCGR (A)              | BovineHD1100009276    |
| DZIP3° (A)             | BovineHD0100015047     | OCLN (A)               | BovineHD2000003255    |
| LHCGR° (A)             | BovineHD1100009276     |                        |                       |
| OCLN (B)               | BovineHD2000003255     |                        |                       |

Gene (DA) = Gene associated with the reproductive trait, in parenthesis the desirable allele reported by Cochran et al. (2013) and Ortega et al. (2015). Gene°, genes that have been reported with overdominance or dominance.

**Table 2.** Top 10 single nucleotide polymorphisms (SNP) associated with heifer and cow conception rate in Braunvieh Cattle.

| Cow conception rate    |                        | Heifer conception rate |                        |
|------------------------|------------------------|------------------------|------------------------|
| Gene (DA) <sup>1</sup> | SNP                    | Gene (DA) <sup>1</sup> | SNP                    |
| APBB1° (A)             | BovineHD1500013531     | CSNK1E (A)             | Hapmap39945-BTA-75021  |
| BCAS1 (A)              | BovineHD1300023602     | DNAH11 (A)             | BovineHD0400008600     |
| CSNK1E (A)             | Hapmap39945-BTA-75021  |                        | Hapmap38265-BTA-96973  |
| DNAH11 (A)             | BovineHD0400008600     | DSC2° (A)              | BovineHD2400007184     |
|                        | Hapmap38265-BTA-96973  |                        | Hapmap52659-ss46526625 |
| DSC2 (B)               | BovineHD2400007184     | DZIP3° (A)             | BovineHD0100015047     |
|                        | Hapmap52659-ss46526625 | FYB1 (B)               | BTB-00778141           |
| GOLGA4° (B)            | BTB-01184624           |                        | BovineHD2000010086     |
| IBSP (A)               | BovineHD0600010277     | GOLGA4 (B)             | BTB-01184624           |
| OCLN (B)               | BovineHD2000003255     | IBSP (A)               | BovineHD0600010277     |

Gene (DA) = Gene associated with the reproductive trait, in parenthesis the desirable allele reported by Cochran et al. (2013) and Ortega et al. (2015). Gene°, genes that have been reported with overdominance or dominance.

**Table 3.** Genetic diversity indices by marker associated with reproductive traits in the Mexican Braunvieh cattle population: observed (Ho) and expected (He) heterozygosity and their standard deviation (SD).

| Marker name (locus)     | He $\pm$ SD       | Ho $\pm$ SD       |
|-------------------------|-------------------|-------------------|
| ARS-BFGL-NGS-4967       | 0.466 $\pm$ 0.132 | 0.477 $\pm$ 0.100 |
| ARS-BFGL-NGS-5623       | 0.474 $\pm$ 0.135 | 0.503 $\pm$ 0.141 |
| BovineHD0100015047      | 0.457 $\pm$ 0.130 | 0.420 $\pm$ 0.115 |
| BovineHD0400008600      | 0.497 $\pm$ 0.141 | 0.493 $\pm$ 0.110 |
| BovineHD0600010277      | 0.502 $\pm$ 0.144 | 0.487 $\pm$ 0.099 |
| BovineHD1000003029      | 0.369 $\pm$ 0.104 | 0.313 $\pm$ 0.577 |
| BovineHD1100009276      | 0.490 $\pm$ 0.141 | 0.500 $\pm$ 0.140 |
| BovineHD1300023602      | 0.493 $\pm$ 0.141 | 0.427 $\pm$ 0.135 |
| BovineHD1500013531      | 0.500 $\pm$ 0.144 | 0.497 $\pm$ 0.100 |
| BovineHD1500021362      | 0.457 $\pm$ 0.129 | 0.527 $\pm$ 0.142 |
| BovineHD2000003255      | 0.476 $\pm$ 0.100 | 0.453 $\pm$ 0.097 |
| BovineHD2000010086      | 0.501 $\pm$ 0.134 | 0.520 $\pm$ 0.140 |
| BovineHD2400007183      | 0.452 $\pm$ 0.100 | 0.420 $\pm$ 0.105 |
| BovineHD2400007184      | 0.487 $\pm$ 0.101 | 0.453 $\pm$ 0.089 |
| BTB-00778141            | 0.498 $\pm$ 0.110 | 0.510 $\pm$ 0.142 |
| BTB-01184624            | 0.490 $\pm$ 0.098 | 0.460 $\pm$ 0.151 |
| Hapmap38265-BTA-96973   | 0.499 $\pm$ 0.100 | 0.487 $\pm$ 0.100 |
| Hapmap39945-BTA-75021   | 0.499 $\pm$ 0.110 | 0.500 $\pm$ 0.139 |
| Hapmap45323-BTA-90907   | 0.419 $\pm$ 0.100 | 0.433 $\pm$ 0.156 |
| Hapmap52659-ss46526625  | 0.493 $\pm$ 0.123 | 0.487 $\pm$ 0.115 |
| Hapmap60475-rs29022896  | 0.488 $\pm$ 0.121 | 0.540 $\pm$ 0.134 |
| Average for the 52 loci | 0.470 $\pm$ 0.122 | 0.420 $\pm$ 0.087 |



**Table 4.** Clustering of 150 individuals (only five representative animals per group are shown) for reproductive traits according to the K-means' method, and proportion tests for the desirable genotypes and alleles of representative animals in each group, in a Mexican population of Braunvieh Cattle.

| Trait <sup>1</sup> | Group | NA <sup>2</sup> | Representative animals              | Representative genotype | DGP <sup>3</sup>   | DAP <sup>4</sup>   |
|--------------------|-------|-----------------|-------------------------------------|-------------------------|--------------------|--------------------|
| PR                 | I     | 62              | 2, 23, 31, 135, 141 <sup>®</sup>    | 2 2 2 0 1 0 0 1 1 0 2 1 | 0.33 <sup>a</sup>  | 0.54 <sup>a</sup>  |
|                    | II    | 88              | 1, 24, 50, 74 <sup>®</sup> , 75     | 1 1 1 0 1 1 1 0 0 1 1 2 | 0.33 <sup>a</sup>  | 0.63 <sup>a</sup>  |
| PL                 | I     | 54              | 21, 33, 48, 50, 132 <sup>®</sup>    | 0 0 0 1 1 1 1 1 1 0 1   | 0.20 <sup>ab</sup> | 0.55 <sup>ab</sup> |
|                    | II    | 56              | 26, 35, 62, 104, 123 <sup>®</sup>   | 2 2 0 1 2 0 1 1 0 1     | 0.33 <sup>a</sup>  | 0.65 <sup>a</sup>  |
|                    | III   | 40              | 61, 78, 113, 133 <sup>®</sup> , 138 | 2 1 0 1 2 1 2 0 1 0     | 0.10 <sup>b</sup>  | 0.45 <sup>b</sup>  |
| CCR                | I     | 66              | 30, 31, 65, 104, 135 <sup>®</sup>   | 1 1 0 2 2 1 1 0 1 0     | 0.20 <sup>a</sup>  | 0.50 <sup>a</sup>  |
|                    | II    | 84              | 58, 75, 90, 112 <sup>®</sup> , 134  | 2 2 1 1 1 2 0 2 0 0     | 0.33 <sup>a</sup>  | 0.50 <sup>a</sup>  |
| HCR                | I     | 96              | 13, 54 <sup>®</sup> , 75, 91, 129   | 2 1 1 1 2 1 1 1 1 1     | 0.20 <sup>a</sup>  | 0.55 <sup>a</sup>  |
|                    | II    | 54              | 3, 93 <sup>®</sup> , 104, 141, 146  | 1 1 2 2 1 1 1 1 0 2     | 0.20 <sup>a</sup>  | 0.45 <sup>a</sup>  |

<sup>1</sup>PR = Pregnancy rate. PL = Productive life. CCR = Cow conception rate. HCR = Heifer conception rate.

<sup>2</sup>NA = Number of animals. <sup>3</sup>DGP = Desirable genotype proportion. <sup>4</sup>DAP = Desirable allele proportion.

Genotype coding: AA=0, AB=1, BB=2. The order of the loci in representative genotype is the same as in Tables 1 and 2, for each trait. Animal<sup>®</sup> is the representative animal (the closest to the centroid). The null hypothesis of non-significant differences between proportions was tested using X<sup>2</sup> and 0.05 significance level; proportions with the same letter are not significantly different.

**Table 5.**

Proportion tests using  $X^2$  test between K-means groups by reproductive trait, for frequencies of desirable alleles per gene in a Mexican Braunvieh population.

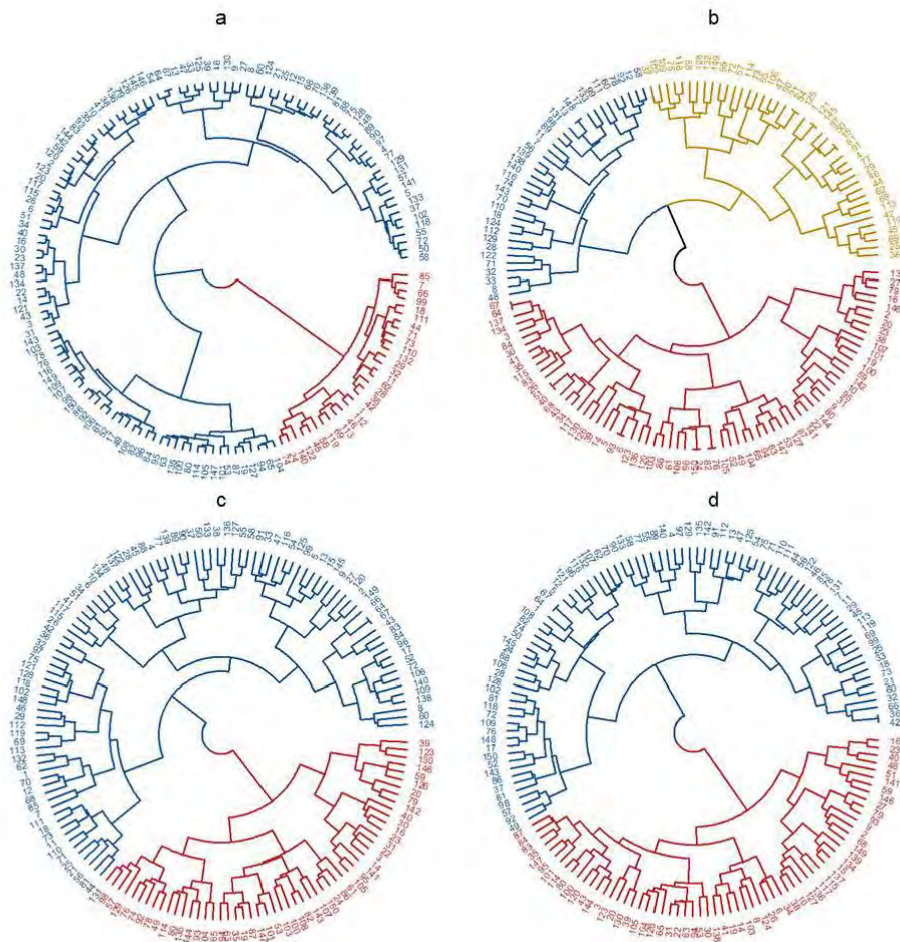
| Genes    | PRG               |                   | PLG               |                   |                   | CCRG              |                   | HCRG              |                   |
|----------|-------------------|-------------------|-------------------|-------------------|-------------------|-------------------|-------------------|-------------------|-------------------|
|          | I                 | II                | I                 | II                | III               | I                 | II                | I                 | II                |
| APBB1    | 0.42 <sup>a</sup> | 0.55 <sup>a</sup> |                   |                   |                   | 0.40 <sup>b</sup> | 0.61 <sup>a</sup> |                   |                   |
| AP3B1    |                   |                   | 0.61 <sup>a</sup> | 0.51 <sup>a</sup> | 0.52 <sup>a</sup> |                   |                   |                   |                   |
| BCAS1    | 0.42 <sup>a</sup> | 0.44 <sup>a</sup> |                   |                   |                   | 0.42 <sup>a</sup> | 0.45 <sup>a</sup> |                   |                   |
| BOLA-DMB | 0.40 <sup>a</sup> | 0.59 <sup>a</sup> |                   |                   |                   |                   |                   |                   |                   |
| CSNK1E   | 0.43 <sup>a</sup> | 0.49 <sup>a</sup> |                   |                   |                   | 0.60 <sup>a</sup> | 0.46 <sup>a</sup> | 0.52 <sup>a</sup> | 0.56 <sup>a</sup> |
| DNAH11   | 0.51 <sup>a</sup> | 0.50 <sup>a</sup> |                   |                   |                   | 0.55 <sup>a</sup> | 0.45 <sup>a</sup> | 0.50 <sup>a</sup> | 0.30 <sup>b</sup> |
| DSC2     | 0.25 <sup>b</sup> | 0.63 <sup>a</sup> | 0.19 <sup>c</sup> | 0.69 <sup>b</sup> | 0.88 <sup>a</sup> | 0.73 <sup>a</sup> | 0.29 <sup>b</sup> | 0.65 <sup>a</sup> | 0.00 <sup>b</sup> |
| DZIP3    | 0.29 <sup>a</sup> | 0.39 <sup>a</sup> |                   |                   |                   |                   |                   | 0.38 <sup>a</sup> | 0.3 <sup>a</sup>  |
| FSHR     |                   |                   | 0.33 <sup>b</sup> | 0.62 <sup>a</sup> | 0.35 <sup>b</sup> |                   |                   |                   |                   |
| FYB1     |                   |                   |                   |                   |                   |                   |                   | 0.54 <sup>a</sup> | 0.45 <sup>a</sup> |
| GOLGA4   |                   |                   |                   |                   |                   | 0.73 <sup>a</sup> | 0.38 <sup>b</sup> | 0.51 <sup>b</sup> | 0.70 <sup>a</sup> |
| HSD17B12 |                   |                   | 0.53 <sup>a</sup> | 0.51 <sup>a</sup> | 0.31 <sup>b</sup> |                   |                   |                   |                   |
| IBSP     |                   |                   | 0.48 <sup>a</sup> | 0.49 <sup>a</sup> | 0.33 <sup>b</sup> | 0.56 <sup>a</sup> | 0.42 <sup>a</sup> | 0.48 <sup>a</sup> | 0.53 <sup>a</sup> |
| LHCGR    | 0.40 <sup>b</sup> | 0.61 <sup>a</sup> | 0.42 <sup>c</sup> | 0.92 <sup>a</sup> | 0.59 <sup>b</sup> |                   |                   |                   |                   |
| OCLN     | 0.99 <sup>a</sup> | 0.38 <sup>b</sup> | 0.32 <sup>b</sup> | 0.21 <sup>b</sup> | 0.52 <sup>a</sup> | 0.52 <sup>b</sup> | 0.73 <sup>a</sup> |                   |                   |
| Average  | 0.46 <sup>a</sup> | 0.51 <sup>a</sup> | 0.41 <sup>b</sup> | 0.57 <sup>a</sup> | 0.50 <sup>a</sup> | 0.56 <sup>a</sup> | 0.47 <sup>a</sup> | 0.51 <sup>a</sup> | 0.41 <sup>a</sup> |

PRG = Pregnancy rate groups. PLG = Productive life groups. CCRG = Cow conception rate groups.

HCRG = Heifer conception rate groups. The null hypothesis established was that there were non-

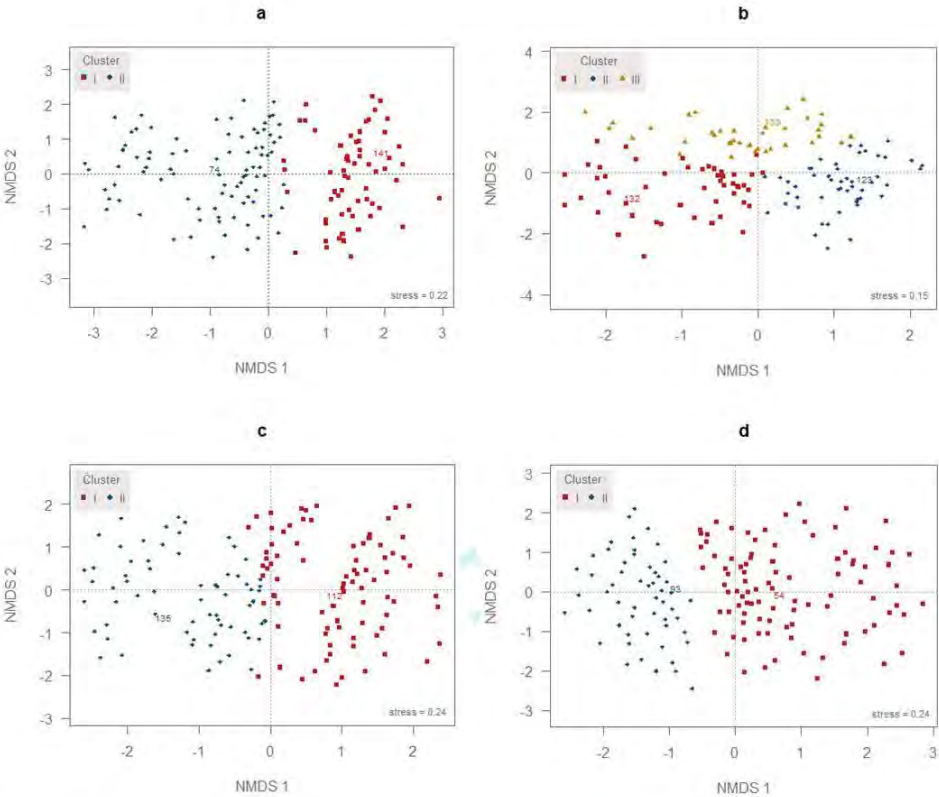
significant differences between proportions, using  $X^2$  test and 0.05 significance level, proportions with the

same letter are not significantly different



**Figure 1.** Dendrograms obtained by Ward's algorithm based on Euclidian distance for Pregnancy Rate (a), Productive Life (b), Cow Conception Rate (c), and Heifer Conception Rate (d) in a Mexican Braunvieh Cattle population.

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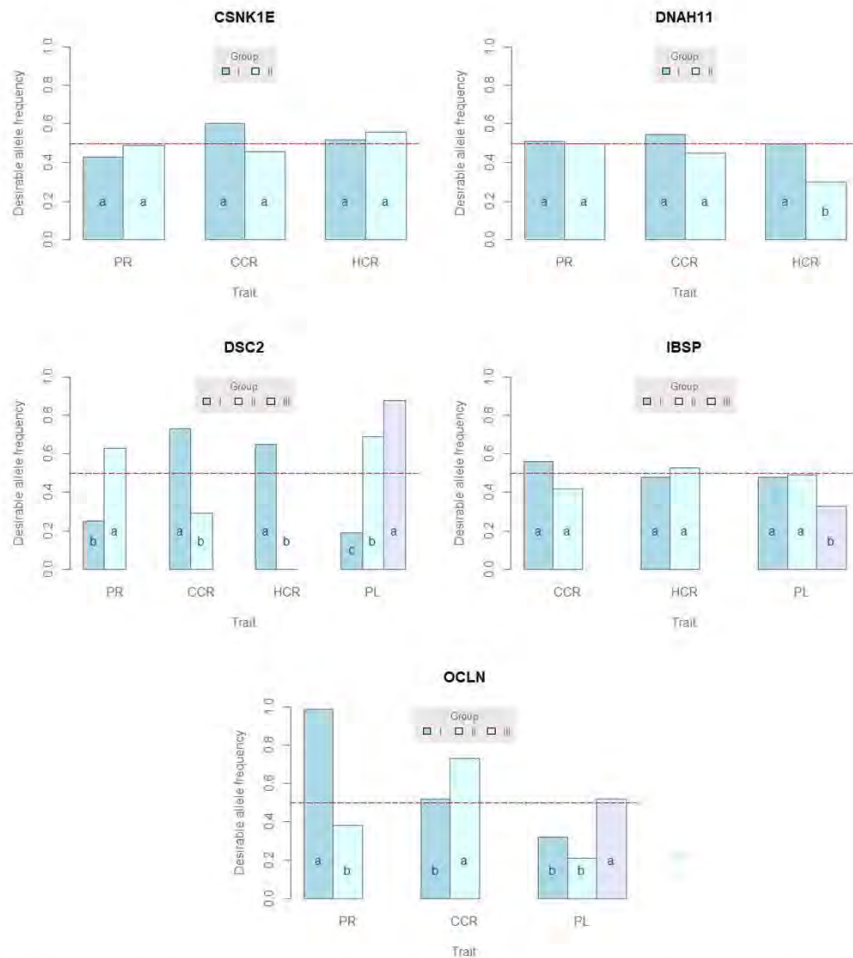


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602 **Figure 2.** Graphic representation of the non-metric multidimensional scaling (NMDS) for  
603 Pregnancy Rate (a), Productive Life (b), Cow Conception Rate (c), and Heifer  
604 Conception Rate (d), using K-means clusters to visualize graphic differences and the  
605 representative animals, in a Mexican Braunvieh Cattle population.

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608 **Figure 3.** Graphic representation of proportion tests using  $\chi^2$  test between K-means on

609 groups classified by reproductive trait for frequencies of desirable alleles of the most

610 important genes in a Mexican Braunvieh population. PR = Pregnancy rate. PL =

611 Productive life. CCR = Cow conception rate. HCR = Heifer conception rate. The null

612 hypothesis established that there were non-significant differences between proportions,

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For Peer Review



# CAPÍTULO 5. GENETIC DIVERSITY AND POPULATION STRUCTURE FOR RESISTANCE AND SUSCEPTIBILITY TO MASTITIS IN BRAUNVIEH CATTLE



Article

## Genetic diversity and population structure for resistance and susceptibility to mastitis in Braunvieh cattle

Mitzilín Zuleica Trujano-Chavez <sup>1</sup>, Reyna Sánchez-Ramos <sup>2</sup>, Paulino Pérez-Rodríguez<sup>3</sup> and Agustín Ruiz-Flores <sup>1,\*</sup>

<sup>1</sup> Posgrado en Producción Animal, Universidad Autónoma Chapingo, Carretera Federal México- Texcoco Km 38.5, 56227, Texcoco, Estado de México, México; zulealizee@gmail.com; aruizf@chapingo.mx

<sup>2</sup> Recursos Genéticos y Productividad; rey\_1014@hotmail.com

<sup>3</sup> Socio Economía Estadística e Informática-Estadística, Colegio de Postgraduados, Carretera Federal México- Texcoco Km 36.5, 56230, Texcoco, Estado de México; perpdgo@gmail.com

\* Correspondence: aruizf@chapingo.mx; Tel.: +52 595 952 1621

**Abstract:** Mastitis is a disease that causes significant economic losses since resistance to mastitis is a difficult trait to improve due to its multifactorial occurrence. Therefore, our objective was to characterize a Mexican Braunvieh cattle population for genetic resistance and susceptibility to mastitis. We used 66 SNP markers for 45 candidate genes in 150 animals. Average heterozygosity was  $0.445 \pm 0.076$ , a value higher than those reported for some European breeds. The inbreeding coefficient was slightly negative for resistance to subclinical ( $-0.058 \pm 0.055$ ) and clinical ( $-0.034 \pm 0.076$ ) mastitis, possibly due to low selection for the immunological candidate genes that influence these traits. The genotypic profiles for the candidate loci per K-means group were obtained, as well as the groups distribution through the graphics of the principal component analysis. The genotypic profiles showed high genetic diversity among groups. Resistance to clinical mastitis had the lowest presence of heterozygous genotypes. Although the percentage of highly inbred animals ( $>50\%$ ) is up to 13.3%, there are highly heterozygous groups in terms of the studied traits, a favorable indicator of the presence of genetic diversity. The results of this study constitute evidence of the genetic potential of the Mexican Braunvieh population to improve mastitis-related traits.

**Keywords:** inbreeding, heterozygosity, candidate genes, principal components analysis

**Citation:** Trujano-Chavez, M.Z.; Sánchez-Ramos, R.; Pérez-Rodríguez, P.; Ruiz-Flores, A. Genetic diversity and population structure for resistance and susceptibility to mastitis in Braunvieh cattle. *Vet. Sci.* **2021**, *8*, x. <https://doi.org/10.3390/vxxxx>

Academic Editor: Firstname Last-name

Received: date

Accepted: date

Published: date

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### 1. Introduction

One of the major problems in dairy units is mastitis [1]. Tropical dual-purpose production systems have the same problem. A prevalence of 60% was reported for mastitis in tropical production units [2]. The current trend seems to focus on milk quality rather than on quantity, and mastitis has been one of the main hurdles [1,3].

Some of the obstacles that traditional genetic selection has faced in bovines are [4]: 1) complexity of quantitative or low-heritability traits, 2) traits measured late in the life of the animals, and 3) traits that are difficult to measure, such as resistance and susceptibility to mastitis.

Resistance to mastitis is a complex trait that is multifactorial in nature and, thus, difficult to improve. The genomic prediction ability for clinical mastitis is low, 0.19 [5], compared to that reported for residual feed intake ( $0.85 \pm 0.08$ ), dry matter intake ( $0.52 \pm 0.04$ ) and feed conversion ratio ( $0.46 \pm 0.05$ ) [6]. The conventional selection scheme for resistance has not been effective against mastitis due to the large number of candidate genes related to several non-immune factors that influence it and challenge its measurement [4, 7–8].

In addition to the control of environmental factors to prevent it at the farm level, genomic methods have been used as an alternative solution to the problem of increasing resistance to mastitis. The aim is to obtain animals that are immune to the main mastitis-

causing pathogens and have qualities that allow them to adapt to environmental factors that cause mastitis [2].

Recently, genotyping of animals has become easy and relatively cheap. A sample of hair or blood will provide thousands of markers [4]. The markers most used for genomic selection are the single polymorphism nucleotides (SNP). Their generalized use has reduced the cost of genotyping to a cost-effective level. Genotyping is widespread, used in both commercial and research systems [9].

Manipulation of genetic variation is key to the development and use of any genetic improvement program for dairy and beef cattle breeds [10]. Genetic diversity has been studied in original Braunvieh and derivative breeds with the aims of disentangling the genetic basis of their adaptation to diverse environments and increasing productive performance. The results have been positive. It was found that populations of this breed adapted to environments different from that of its origin are genetically more diverse than the average of other *Bos taurus* and some *Bos indicus* breeds [11]. Animals living in harsh environmental conditions, such as extreme temperatures, possess greater genetic diversity because facing environmental changes requires versatility [12].

The objective of this study was to determine the genetic diversity and structure of a Mexican Braunvieh population in terms of mastitis resistance and susceptibility, using 1) basic population statistics: inbreeding coefficient, expected and observed heterozygosity, and 2) multivariate statistics: K-means and principal components analysis.

## 2. Materials and Methods

### 2.1. Source of information

Genomic information was obtained using hair samples from 150 animals born between 2001 and 2016 on five farms belonging to the Mexican Association of Braunvieh Purebred Breeders in Eastern, Central, and Western Mexico. The samples were genotyped at GeneSeek (Lincoln, NE, USA, <http://genomics.neogene.com>). The chip used was the Genomic Profile Bovine LD with 50,000 SNP markers.

### 2.2. Genotype quality control

For data edition, quality control was performed prior to the genotypic analyses. The missing genotypes were imputed using the observed allele frequencies. The imputation method consisted of assigning an allele, according to the probability of a possible type of polymorphism for a certain marker in the entire population. Additionally, a 0.05 threshold for the minor allele frequency was considered to discard non-informative markers or those with errors in genotyping.

### 2.3. Statistical analyses

#### 2.3.1. Identification of associated and informative loci

The candidate genes associated with mastitis in *Bos taurus* were identified, according to reports by [13–16] for resistance to mastitis (RM); by [17–18] for susceptibility to mastitis (SM); by [19–21] for resistance to bacterial mastitis (RBM); by [8] for resistance to subclinical mastitis (RSM); and by [22] for resistance to clinical mastitis (RCM).

The position ranked by base pairs were obtained from the Gene Library [23]. With this information, the intragenic loci were searched; the criterion for the loci-gene association was that they should be located within the position rank  $\pm 25$  kb of the gene for the same chromosome. Only the loci available in the genotypic database were included. The analyses considered 455 SNP markers of 204 candidate genes reported in the literature associated with resistance and susceptibility to mastitis.

All the analyses were carried out using the R software, version 4.0.4 [24]. According to the literature consulted, the 10% most informative markers associated with each of the

studied traits were determined using the Shannon index. This index allows identification of the most informative markers [25]. The index is defined as:

$$H = - \sum_{i=0}^2 p_i \log p_i$$

Where  $i = 0, 1, 2$  refers to the genotype (AA = 0, AB = 1, BB = 2) and  $p_i$  is the corresponding allelic frequency. This index was calculated for each of the loci. Empirical distribution was obtained, and the 10% most informative markers were selected. The highest values for  $H$  were associated with the highest diversity for the marker. At the end of filtering, 66 SNP markers of 45 candidate genes were left for further analyses. For better handling of the markers in the graphs, the names of the loci were replaced by consecutive numbers, assigned by alphabetical order to the corresponding candidate gene.

### 2.3.2. Population genetic structure

Expected ( $H_e$ ) and observed ( $H_o$ ) heterozygosity was estimated for all the loci by trait in the study by using the adegenet program [26] available in R [24]. The stats program of R [24] was used to perform the t-test to determine the differences between  $H_e$  and  $H_o$ . The null hypothesis was that there were non-significant differences between them. The p-value 0.01, used to declare significance, and the p-value was corrected with the Bonferroni method considering the total number of loci by trait.

The program pegas [27] through adegenet [26] was used to determine whether the loci under study were in Hardy-Weinberg equilibrium. A hundred thousand replicates were assigned to execute the test procedure through the Monte Carlo method. The null hypothesis established the existence of Hardy-Weinberg equilibrium when p-values were higher than  $0.01/n$  with the Bonferroni correction;  $n$  is the number of loci per trait.

The inbreeding coefficients by individual with respect to the population ( $F_{IT}$ ) were calculated and plotted with adegenet [26]. Additionally, the general mean for  $F_{IT}$  per trait was obtained, along with its respective standard deviation. Finally, a histogram including a nonparametric estimate of the density function by trait was obtained. This was done to visualize the animals with an inbreeding coefficient above 0.5 (the number of animals could be observed in the histogram).

### 2.3.3. Cluster analyses

The differences in the genotypic profile of individuals were identified with two clustering algorithms: the hierarchical clustering using the Ward distance implemented in the stats package and the K-means algorithm also included in the stats package in R and factoextra [28]. For both methods, the 10% of the markers, identified in a previous step, was used.

The optimum number of groups ( $k$ ) to be formed through the hierarchical clustering and K-means was obtained with Silhouette and Elbow of factoextra [28]. A search for  $k$  that allowed equilibrium between a value of the silhouette close to one [29] and the least error sum of squares was carried out.

The Ward algorithm of minimum variance, according to [30], is a divisive hierarchical method that creates mutually exclusive sub-sets. Sub-set members are ultimately similar to each other and different between groups. This promises to be a good method for finding different groups with particular genotypic profiles. The results of the procedure are illustrated in a circular dendrogram that displays the hierarchy of the groups and their similarity to the groups formed with the K-means algorithm.

K-means is a non-hierarchical algorithm that uses a reassignment method based on centroids to form groups. This is the most frequently used method to form groups [31]. It is used in robotics, genomics, and genetic diversity. In our study the groups of K-means were used to identify divergent genotypes among groups of animals and to characterize genetic diversity based on the Principal Components Analysis (PCA).

#### 2.3.4. Principal Components Analysis

The PCA was performed using adegenet package [26] using the sample covariance matrix obtained using the candidate SNP. The objective was to observe the distribution of the groups formed by K-means through a PCA graphic of individuals (1<sup>st</sup> and 2<sup>nd</sup> dimensions) and to reduce the number of markers according to their eigenvalues. With a factor-extra biplot [28], we estimated the correlation and contribution of markers to the 1<sup>st</sup> and 2<sup>nd</sup> dimensions formed by the PCA and determined the 10% of the animals with the highest contribution to the analysis.

### 3. Results and Discussion

#### 3.1. Sample size and power analysis

In México, studies using genomic information from large animal populations are scarce. Usually, the main restriction is the availability of economic resources for animal genotyping. Genetic improvement infrastructure of developing countries usually is less robust than that of developed countries whose genomic databases involve millions of animals [32].

In previous studies carried out with the same population used in our study [33–34], genomic information of 300 animals was considered. For our study only 150 animals with candidate SNPs associated with resistance and susceptibility to mastitis were found.

The sample size used in our study, evaluated with the `pwr.chisq.test` function of the `pwr` package [35], was enough to detect significant differences with a p-value of 0.05 and an effective size of 0.4. The power of 0.87 for the  $\chi^2$  test obtained in the present study is adequate to obtain conclusive, reliable results [36]. In other studies of genetic diversity and structure, smaller sample sizes have been used, and conclusive results have been reached: 6 to 23 [37], 7 to 58 [38], 13 to 38 [39], and 71 to 167 [40].

#### 3.2. Identification of associated and informative loci

Tables 1 to 5 show the markers obtained after filtering with the Shannon Index. Their candidate genes, chromosome, and the number assigned to visualize the results in the graphs are shown. For RM, 19 genes with 22 loci were obtained; for SM, seven genes and 11 loci; for RBM, seven genes and eight markers; for RSM, eight genes and 20 markers; and for RCM, four genes and six markers. This adds up to 66 markers and 45 candidate genes. All loci were in Hardy-Weinberg equilibrium.

#### 3.3. Population genetic structure

Table 6 shows the results for  $H_e$  and  $H_o$ , as well as those of a t-test to determine whether the differences were statistically significant. The rule of decision to not reject the null hypothesis of non-significant differences between  $H_e$  and  $H_o$  was that the p-value for the t-test > p-value for BC. In the last column the  $|H_e - H_o|$  differences can be observed; all were non-significant, with exception of the difference for SM.

The estimates for  $H_o$  obtained in our study for mastitis (average  $0.45 \pm 0.076$ ) are above those obtained with breeds derived from the original Braunvieh, but below those reported for *Bos taurus* breeds with some adaptation to the tropics. In Colombia for Creole cattle, [41] reported  $H_o$  of 0.66; the Costeño con Cuernos breed had the lowest, 0.635, and Casarcño the highest, 0.733. These  $H_o$  estimates are quite different from those reported by [42], who found an average of  $0.35 \pm 0.167$  for  $H_o$  in populations related to Braunvieh.

This suggests that the Mexican Braunvieh population has increased its genetic versatility for the candidate genes studied, with a higher number of heterozygous individuals. Exposure to climates different from their native climate for over a century in the tropical systems of Mexico could have caused a change in  $H_o$ , with respect to original Braunvieh populations [42–43].

**Table 1.** Top 10% single nucleotide polymorphisms (SNP) associated with Resistance to Mastitis (RM) in Braunvieh Cattle.

| Gene           | Chr | Position, pb ( $\pm$ 25 k) | SNP marker name        | LN |
|----------------|-----|----------------------------|------------------------|----|
| ARHGAP10 [15]  | 17  | 9994895-10429767           | ARS-BFGL-NGS-113821    | 1  |
|                |     |                            | BovineHD1700002890     | 2  |
| BDH2 [14]      | 6   | 21737056-21815795          | BovineHD0600006001     | 3  |
| CAPG [13]      | 11  | 49562863-49630835          | Hapmap54495-rs29018810 | 4  |
| CHD5 [13]      | 16  | 47131943-47253618          | BovineHD1600013051     | 5  |
| CST6 [14]      | 29  | 44076083-44127573          | BovineHD2900013196     | 6  |
| ELMO1 [13]     | 4   | 59842686-60475511          | BovineHD0400016236     | 7  |
| FBL [15]       | 18  | 49363469-49425788          | ARS-BFGL-NGS-113564    | 8  |
| FOCAD [13]     | 8   | 23394437-23750228          | BovineHD0800007122     | 9  |
| IMMP2L [13]    | 4   | 56736178-57747698          | BovineHD0400015680     | 10 |
| LOC510112 [15] | 10  | 27914007-27964951          | BovineHD1000009188     | 11 |
|                |     |                            | BovineHD1000009179     | 12 |
| MBL2 [16]      | 26  | 6306933-6362539            | BovineHD2600001441     | 13 |
|                |     |                            | BovineHD2600001446     | 14 |
| MYO1E [13]     | 10  | 50932729-51205381          | BovineHD1000015285     | 15 |
| PON1 [13]      | 4   | 12517349-12601241          | ARS-BFGL-NGS-48351     | 16 |
| PPP3CA [13]    | 6   | 23419690-23795287          | BovineHD0600006555     | 17 |
| ST7 [13]       | 4   | 51162819-51481394          | BovineHD0400014185     | 18 |
| SULF2 [14]     | 13  | 76130676-76307180          | BovineHD1300022072     | 19 |
| TBC1D8 [13]    | 11  | 5953170-6144340            | BovineHD1100002208     | 20 |
| TBCK [13]      | 6   | 18840254-19099244          | BovineHD0600005226     | 21 |
| ZNFX1 [14]     | 13  | 77323734-77400021          | BovineHD1300022388     | 22 |

Chr = Chromosome. Position, pb ( $\pm$  25 k) = Candidate gene position in base pairs reported in [23], for *Bos taurus*  $\pm$  25,000 base pairs. LN = Locus Number assigned in graphs.

On the other hand, the non-significant differences between He and Ho for most of the traits (Table 6) is similar to the results obtained in other studies, where neither the breed nor the environment modified the non-rejection of the null hypothesis, He=Ho for  $\chi^2$  [41-44].

As a graphic resource, Figure 1 shows that the absolute differences at the locus level are small, as in the overall result. The exception to this was SM whose joint trend of the loci was to be over 0.03. Thus, it is the only trait with significant differences. In the bar-plots for the rest of the traits, there were loci that were both close to zero and above 0.05, but the overall estimate is not enough to declare evidence for a significant difference.

The estimate for Fr obtained coincides with those observed in semi-specialized systems of production. The overall average for Fr was  $0.017 \pm 0.043$  for all the traits associated with mastitis. This value is similar to those found for Sahiwal ( $0.013 \pm 0.109$ ), Gyr ( $0.013 \pm 0.106$ ), and Guernsey ( $0.02 \pm 0.217$ ) [45]. In contrast, the Fr for the tropical breeds Landim, Angole, and Tete [46] was 70% higher than that found in our study. Furthermore, for Creole Colombian breeds the average was 0.09 [41]. The first group of breeds was maintained in semi-specialized systems [45], while Landim, Angole, Tete, and the Colombian Creole are non-specialized breeds. This difference could explain why smaller breeds without adequate genetic control present higher inbreeding coefficients.

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**Table 2.** Top 10% single nucleotide polymorphisms (SNP) associated with Susceptibility to Mastitis (SM) in Braunvieh Cattle.

| Gene          | Chr | Position, pb ( $\pm$ 25 k) | SNP marker name       | LN |
|---------------|-----|----------------------------|-----------------------|----|
| ANKRD33B [17] | 20  | 62576901-62728565          | ARS-BFGL-NGS-112060   | 1  |
| CTNND2 [17]   | 20  | 61189491-62339843          | BovineHD2000017160    | 2  |
|               |     |                            | BovineHD2000017153    | 3  |
|               |     |                            | BTB-00791947          | 4  |
| GRIA3 [18]    | 10  | 7542786-7901396            | ARS-BFGL-NGS-87466    | 5  |
| ILDR2 [18]    | 3   | 1878471-1997329            | Hapmap42630-BTA-67480 | 6  |
| ITPK1 [18]    | 21  | 57564349-57778522          | BovineHD2100016547    | 7  |
|               |     |                            | BovineHD2100016592    | 8  |
|               |     |                            | BovineHD2100016552    | 9  |
| SIDT1 [18]    | 1   | 58193233-58355060          | BovineHD0100016494    | 10 |
| TBXAS1 [18]   | 4   | 103270503-103494001        | BovineHD0400029111    | 11 |

Chr = Chromosome. Position, pb ( $\pm$  25 k) = Candidate gene position in base pairs reported in [23] for *Bos taurus*  $\pm$  25,000 base pairs. LN = Locus Number assigned in graphs.

**Table 3.** Top 10% single nucleotide polymorphisms (SNP) associated with Resistance to Bacterial Mastitis (RBM) in Braunvieh Cattle.

| Gene                       | Chr | Position, pb ( $\pm$ 25 k) | SNP marker name     | LN |
|----------------------------|-----|----------------------------|---------------------|----|
| ECHDC1 <sup>1</sup> [21]   | 9   | 23922553-24025911          | BovineHD0900006460  | 1  |
| GNAI1 <sup>2</sup> [19]    | 4   | 40930829-41086584          | ARS-BFGL-NGS-110306 | 2  |
| IL1A <sup>3</sup> [20]     | 11  | 46457553-46518533          | BovineHD1100013546  | 3  |
| IL1RN <sup>2, 3</sup> [19] | 11  | 46790914-46862407          | ARS-BFGL-NGS-113289 | 4  |
|                            | 11  | 46790914-46862408          | ARS-BFGL-NGS-113289 | 4  |
| ITGA4 <sup>2</sup> [19]    | 2   | 15058580-15200364          | BovineHD0200004269  | 5  |
| ITGB3 <sup>2</sup> [19]    | 19  | 46305531-46418474          | UA-IFASA-8333       | 6  |
| NRG1 <sup>1</sup> [21]     | 27  | 28396076-28680095          | BovineHD2700007962  | 7  |

Superscripts indicate resistance to <sup>1</sup> *S. aureus*, <sup>2</sup> *S. agalactiae* and <sup>3</sup> *E. coli*. Chr = Chromosome. Position, pb ( $\pm$  25 k) = Candidate gene position in base pairs reported in [23] for *Bos taurus*  $\pm$  25,000 base pairs. LN = Locus Number assigned in graphs.

Figure 2 shows that, for traits RM, SM, and RBM, most of the loci had positive *F*<sub>IR</sub>. In contrast, the loci for RSM and RCM are mostly negative; that is, there are no traces of inbreeding. This could be due to the specificity of the last two traits, where the candidate genes involve DNA segments that have undergone null direct selection.

The genes whose symbol starts with CXCL are responsible for the immune response [47]. In our study, for RCM, 67% of the SNP markers belong to the genes CXCL1 and CXCL8. This explains the low *F*<sub>IR</sub> values for the trait. This is because current improvement programs for the Braunvieh population under study have no selection criteria based on resistance to mastitis.

For RSM, 40% of the markers belong to EDN2 and HIVEP3, candidate genes associated with immune processes, such as intracellular and sequential signaling of immunoglobulin receptors. Because of their immunologic nature, these genes have not been directly selected for; however, at the farm level there is a rising concern for improving their animals for genes associated with diseases that could be prevented, such as paratuberculosis and clinic mastitis [8, 48].



**Table 4.** Top 10% single nucleotide polymorphisms (SNP) associated with Resistance to Subclinc Mastitis (RSM) in Braunvieh Cattle.

| Gene     | Chr | Position, pb ( $\pm$ 25 k) | SNP marker name        | LN |
|----------|-----|----------------------------|------------------------|----|
| BMPRI1B  | 6   | 29346363-29845366          | BovineHD0600008191     | 1  |
|          |     |                            | BovineHD0600008283     | 2  |
|          |     |                            | BovineHD0600008292     | 3  |
| EDN2     | 3   | 104654329-104730954        | BovineHD0300029984     | 4  |
|          |     |                            | BovineHD0300029996     | 5  |
| GUCA2A   | 3   | 103990493-104056213        | BTA-98478-no-rs        | 6  |
| HEYL     | 3   | 106417713-106483224        | BTB-00148619           | 7  |
| HIVEP3   | 3   | 104084087-104699365        | ARS-BFGL-NGS-35125     | 8  |
|          |     |                            | BovineHD0300029904     | 9  |
|          |     |                            | BovineHD0300029925     | 10 |
|          |     |                            | BovineHD0300029955     | 11 |
|          |     |                            | BTB-01612988           | 12 |
|          |     |                            | Hapmap48983-BTA-100103 | 13 |
| MACF1    | 3   | 106564750-106955676        | ARS-BFGL-NGS-38199     | 14 |
|          |     |                            | BovineHD0300030641     | 15 |
|          |     |                            | BovineHD0300030658     | 16 |
|          |     |                            | BovineHD0300030677     | 17 |
|          |     |                            | BovineHD0300030702     | 18 |
| MFSD2A   | 3   | 106097951-106160784        | BovineHD0300030435     | 19 |
| SH3PXD2A | 26  | 24136301-24435553          | BovineHD2600006239     | 20 |

Chr = Chromosome. Position, pb ( $\pm$  25 k) = Candidate gene position in base pairs reported in [23] for *Bos taurus*  $\pm$  25,000 base pairs. LN = Locus Number assigned in graphs. Candidate genes reported by [8].

**Table 5.** Top 10% single nucleotide polymorphisms (SNP) associated with Resistance to Clinical Mastitis (RCM) in Braunvieh Cattle.

| Gene  | Chr | Position, pb ( $\pm$ 25 k) | SNP marker name    | LN |
|-------|-----|----------------------------|--------------------|----|
| CXCL1 | 6   | 89047989-89100128          | BovineHD0600024410 | 1  |
| CXCL8 | 6   | 88785817-88839572          | ARS-BFGL-NGS-17376 | 2  |
|       |     |                            | BovineHD0600024315 | 3  |
|       |     |                            | BovineHD0600024328 | 4  |
| SEL1L | 10  | 92733415-92848117          | BovineHD1000026808 | 5  |
| STAT4 | 2   | 79543834-79730325          | BovineHD0200022927 | 6  |

Chr = Chromosome. Position, pb ( $\pm$  25 k) = Candidate gene position in base pairs reported in [23] for *Bos taurus*  $\pm$  25,000 base pairs. LN = Locus Number assigned in graphs. Candidate genes reported by [22].

The distribution of animals by their Fr value can be observed in Figure 3. In general, the animals fall in the range of 0 to 0.3 Fr. Fr=50% is equivalent to the value of an inbred animal produced by two progenitors with 100% genetic relationship. Our animals varied from 1.3% to 13.3%. The trait with the highest number of highly inbred animals was SM, while RSM had the lowest.

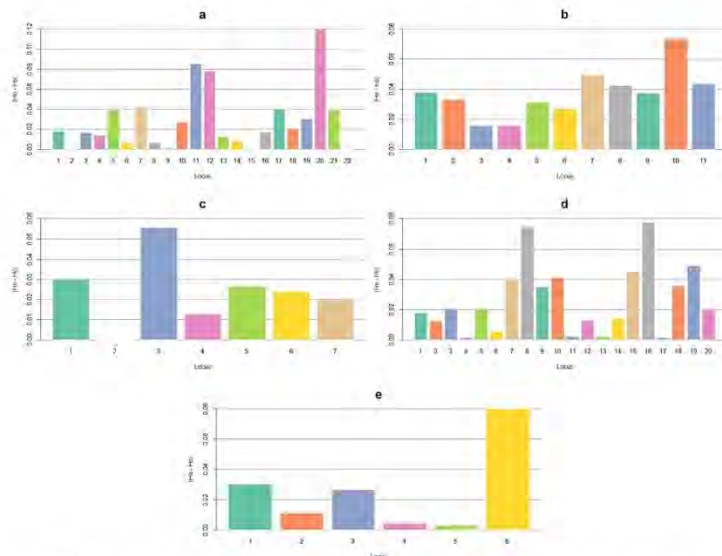
Although up to 13.3% of animals have non-desirable Fr values, the estimate obtained in our study ( $0.017 \pm 0.043$ ) is similar to that reported for other cattle breeds in specialized

and semi-specialized systems of production. In those systems, although some animals had high inbreeding coefficients, the average  $F_{IT}$  is lower than those reported by [49] for *Bos taurus* breeds (0.071), Brown Swiss (0.071), Braunvieh (0.059), Original Braunvieh (0.023), Holstein (0.057), and Simmental (0.028).

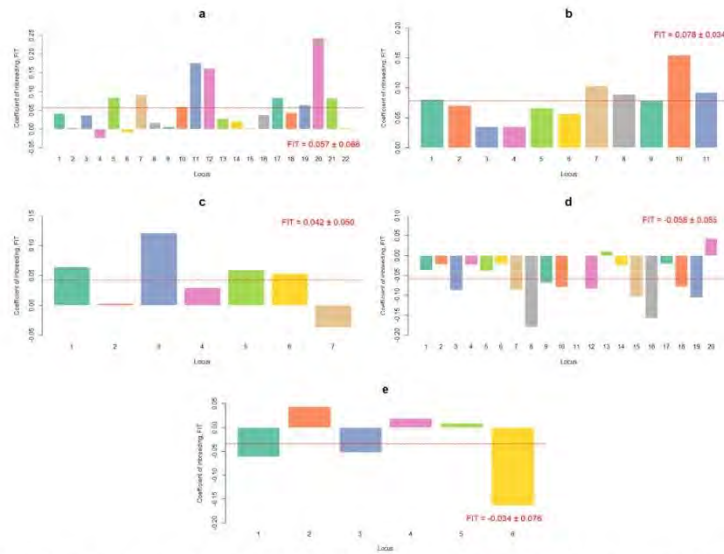
**Table 6.** Heterozygosity and t-test results by trait associated with mastitis, for a Mexican Braunvieh Cattle population.

| Trait                              | He $\pm$ SD       | Ho $\pm$ SD       | NL | p-value BC           | p-value t-test       | He-Ho  |
|------------------------------------|-------------------|-------------------|----|----------------------|----------------------|--------|
| Resistance to Mastitis             | 0.495 $\pm$ 0.006 | 0.468 $\pm$ 0.034 | 22 | 4.5 $\times 10^{-4}$ | 5.5 $\times 10^{-4}$ | 0.027  |
| Susceptibility to Mastitis         | 0.495 $\pm$ 0.004 | 0.458 $\pm$ 0.019 | 11 | 9.1 $\times 10^{-4}$ | 1.0 $\times 10^{-5}$ | 0.037* |
| Resistance to Bacterial Mastitis   | 0.488 $\pm$ 0.010 | 0.470 $\pm$ 0.031 | 7  | 1.4 $\times 10^{-3}$ | 4.4 $\times 10^{-2}$ | 0.018  |
| Resistance to Subclinical Mastitis | 0.379 $\pm$ 0.153 | 0.403 $\pm$ 0.164 | 20 | 5.0 $\times 10^{-4}$ | 9.9 $\times 10^{-1}$ | -0.024 |
| Resistance to Clinical Mastitis    | 0.406 $\pm$ 0.105 | 0.426 $\pm$ 0.130 | 6  | 1.7 $\times 10^{-3}$ | 8.9 $\times 10^{-1}$ | -0.020 |

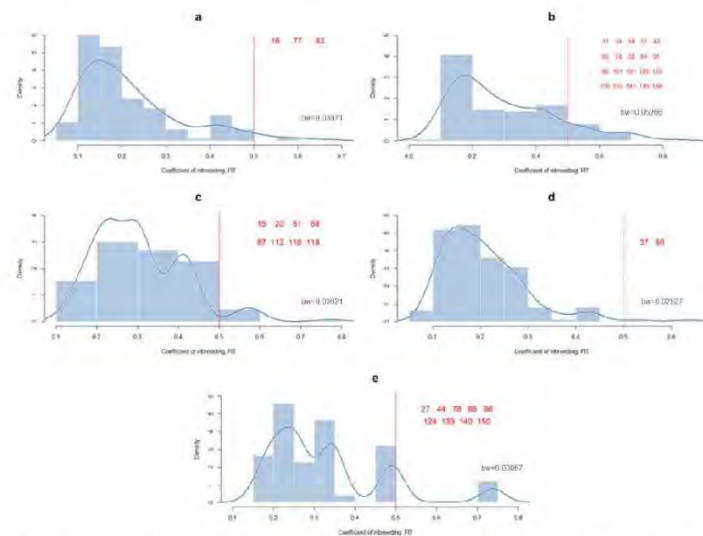
He  $\pm$  SD = Heterozygosity expected  $\pm$  Standard Deviation. Ho  $\pm$  SD = Heterozygosity observed  $\pm$  Standard Deviation. NL = Number of loci. p-value BC = p-value with Bonferroni correction (0.01/NL). p-value t-test = p-value obtained by the t-test, where the null hypothesis established that the differences between He and Ho from zero were non-significant, at 0.01/NL significance level. He-Ho = Differences between heterozygosity values (expected - observed), values\* indicate statistically significant difference.



**Figure 1.** Absolute differences by locus of expected heterozygosity (He) minus observed (Ho) for resistance (a) and susceptibility (b) to mastitis, resistance to bacterial mastitis (c), subclinical mastitis (d) and clinical mastitis (e) in a Mexican Braunvieh cattle population. |He-Ho| = absolute difference between He and Ho. Note: see the names of each locus by trait in Tables 1 to 5.



**Figure 2.** Coefficients of inbreeding ( $F_{IT}$ ) per locus and trait for resistance (a) and susceptibility (b) to mastitis, resistance to bacterial mastitis (c), subclinical mastitis (d) and clinical mastitis (e) in a Mexican Braunvieh cattle population. The mean of  $F_{IT}$  and its standard deviation are shown in red. The red dotted line indicates the mean of  $F_{IT}$  for all the loci. Note: see the names of each locus by trait in Tables 1 to 5.



**Figure 3.** Histogram and density plot (cosine Kernel Function) of the coefficient of inbreeding ( $F_{IT}$ ) for resistance (a) and susceptibility (b) to mastitis, resistance to bacterial mastitis (c), subclinical mastitis (d) and clinical mastitis (e) in a Mexican Braunvieh cattle population. bw= Bandwidth. Animals with a  $F_{IT}$  greater than 0.5 are shown in red on each plot.

### 3.4. Clustering using hierarchical methods and K-means algorithms

The optimum number of  $k$  to equilibrate the silhouette width and the means of squared errors for both algorithms converged at the same number of clusters for each trait. However, better values for silhouette were found by grouping with the K-means algorithm: RM, 0.08; RS, 0.17; RBM, 0.17; RSM, 0.12; and RCM, 0.39. For the first four traits, the silhouette values were low, but in all cases, they were negative, a good indicator of the suitability of the method. The RCM silhouette was the widest.

The average proportion of variability among groups relative to the total variability was obtained. The estimates found were RM, 34.9%; RS, 65.5%; RBM, 66.9%; RSM, 54.7%; and RCM, 77.3%. It should be noted that the RCM values for variability and silhouette were the best of all the traits. This could be due to the small number of markers of the trait (6), which were highly informative as well.

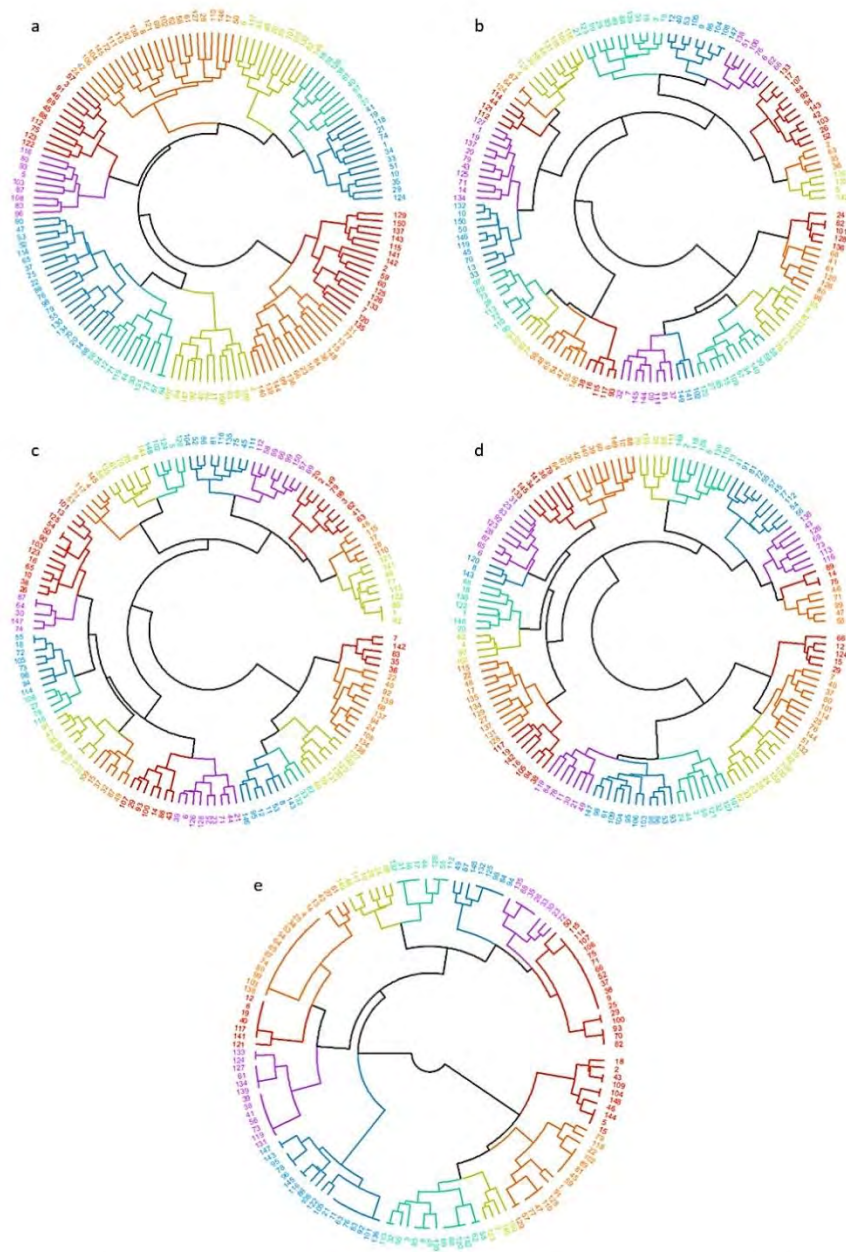
The circular dendrograms of Figure 4 were generated to visualize the groups formed by the Ward hierarchical method. The groups created with this algorithm were very similar to those created with K-means. The  $k$  number, the number of individuals per group, and the representative animals together with their genotypic profile can be seen in Figure 5. The representative animal is the animal closest to the centroid of its cluster, and its genotypic profile was obtained to visualize the main differences among group patterns.

Ho and F<sub>IT</sub> are not related because, unlike inbreeding, heterozygosity is not correlated with the markers [50], even though Ho values close to 0.50 were found for all the traits. In the genotypic profiles, it can be observed that there are groups of animals with low frequency of genotype AB, Figure 5. The traits with the lowest values for F<sub>IT</sub> present high numbers of homozygous animals. This could be explained by the low genetic relationship among the animals for these loci, that is, genotypes AA or BB for the same locus, as seen in Table 6. One of these traits (SM) presents a significant H<sub>e</sub>-Ho difference. This means a decrease in the expected value for heterozygosity which will be reflected in a greater number of homozygous individuals.

The trait with the greatest number of heterozygous animals was RM. Groups III, VII, VIII, and XI possess most of their loci with genotype AB. RM is multifactorial, and therefore, it includes candidate genes associated with other productive, reproductive, and adaptive traits. This will cause high genetic diversity, in contrast with the traits RSM and RCM, which are influenced by immunological candidate genes of specific order whose diversity is limited. According to [51], these gene types tend to be found in homozygous genotypes in wildlife, given that AB genotypes are associated with proteinic abnormalities that increase their susceptibility to pathogenic diseases.

For SM there are well defined groups of animals whose loci are mostly heterozygous. Groups II, IV, V, XV, and XVII have at least 55% of their loci with genotype AB. The groups with fewest heterozygous individuals were III, XVIII, XIX, and XX with up to 18% AB genotypes. There is high genetic diversity in the population for the trait as was observed for RM; the reason could be the presence of genes influencing traits that are indirectly related with mastitis, and not just loci affecting the immune system [47,51].

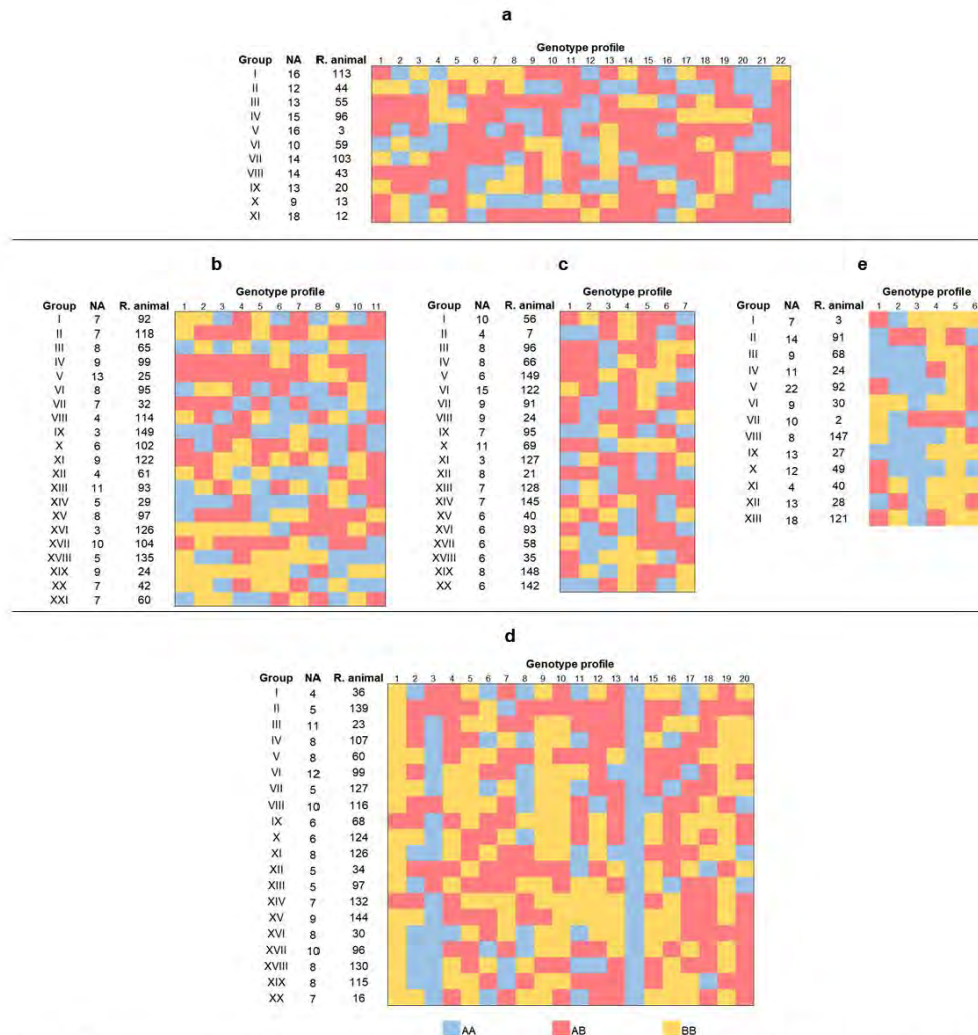
The loci of the groups formed by the algorithm K-means for RBM are from 29% (groups II, VI, IX, and XX) to 57% (groups I, III, VIII, and XII) heterozygous. Diversity was high because only 33% of the genes were directly related with immune functions in the animal, genes IL1A and IL1RN, while the rest was related to metabolic processes [19-21].



**Figure 4.** Circular dendrograms obtained by Ward's algorithm based on Euclidian distance for resistance (a) and susceptibility (b) to mastitis, resistance to bacterial mastitis (c), subclinical mastitis (d) and clinical mastitis (e) in a Mexican Braunvieh cattle population.

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**Figure 5.** Clustering results for 150 individuals (only one representative animal per group is shown) for resistance (a) and susceptibility (b) to mastitis, resistance to bacterial mastitis (c), subclinical mastitis (d) and clinical mastitis (e) in a Mexican Braunvieh cattle population. NA = Number of animals in the group. R. animal = representative animal (the closest to the centroid). Note: the names of each locus by trait are indicated in Tables 1 to 5.

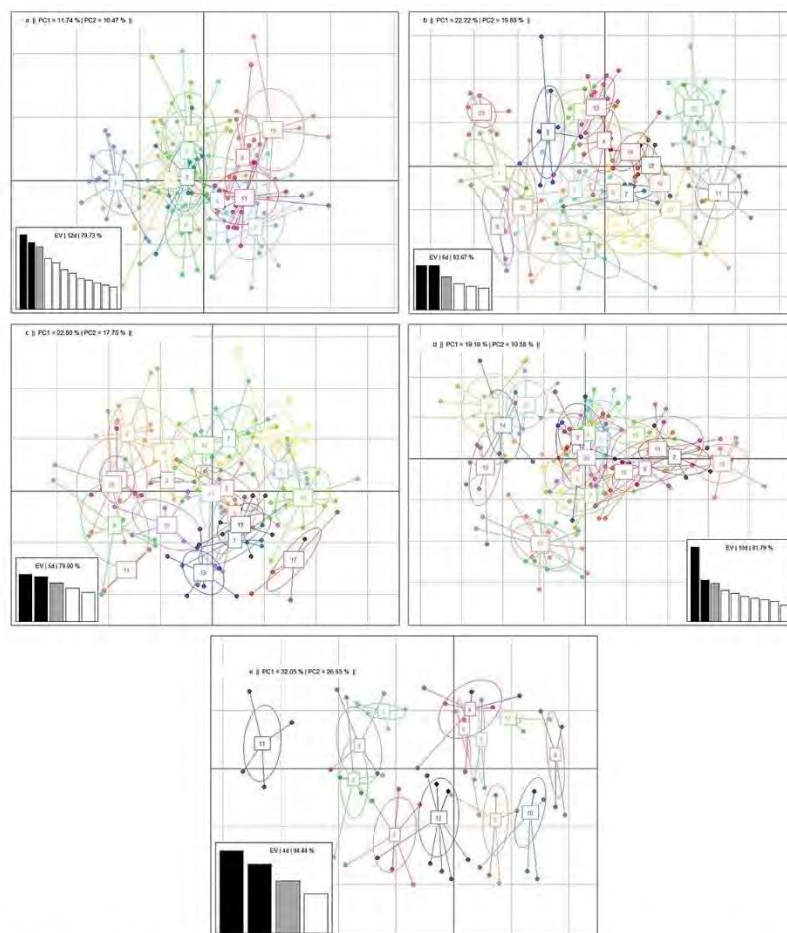
### 3.5. Principal Components Analysis (PCA)

The PCA was useful to observe the genetic diversity in the groups of animals generated with K-means. The first two dimensions of the PCA explained 22.21% to 58.9% of the variation found in the markers associated with mastitis, Figure 6. These results were higher than those reported in similar studies [37,38]. These authors found that 10.9% and 8.79%, respectively, of the variation was explained by the first two dimensions.

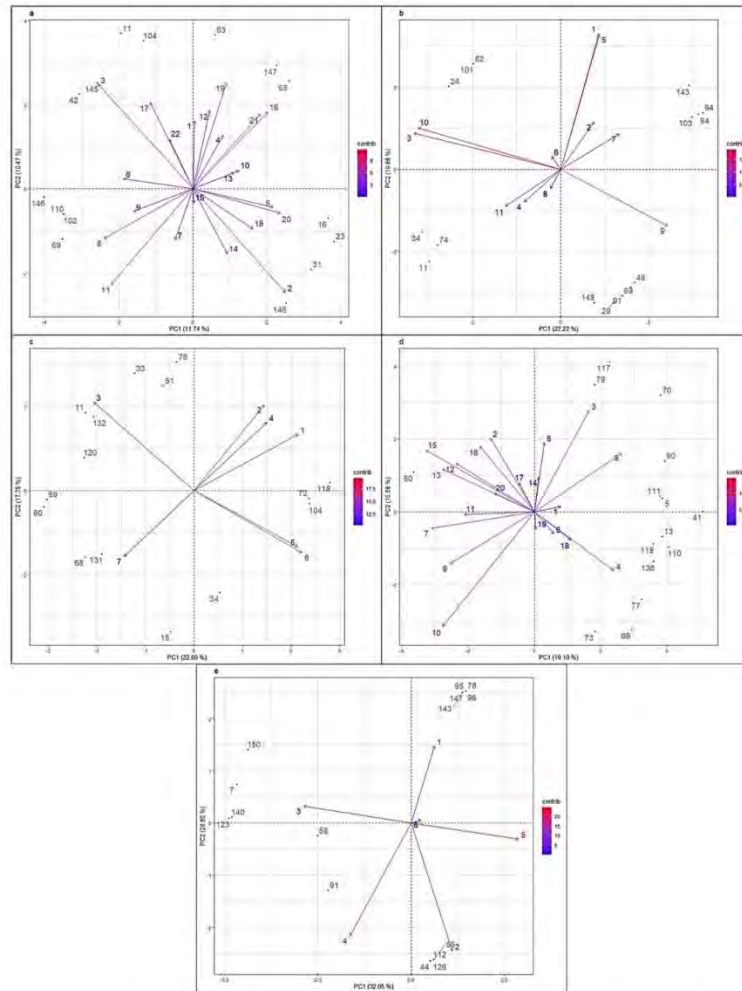


For traits RM and RSM, which possess a larger number of the loci under study, the number of eigenvalues considered to reach a minimum of 80% of variability is higher than that for the rest of the traits. However, the first two dimensions possess a high percentage of variation, evident in the graphic representation of the PCA, where the K-means groups are clearly differentiated, particularly for the RCM graph.

For RM, the genes ARHGAP10 and BDH2 (SNPs 2 and 3, respectively) contributed the most to the analysis; however, the relationship between them is negative (Figure 7). This means the genotypes for the genes in an animal are present as AA and in another as BB, or vice versa. Between these genes there was no known linkage or genetic correlation, given that they were in different chromosomes and affect unrelated traits. ARHGAP10 is associated with intramuscular fat formation [52] and BDH2 with feed intake [53].



**Figure 6.** Graphic representation of the Principal Components Analysis: K-means' grouping of individuals showing eigenvalues (EV) for the dimensions (d) whose sum of variability is around 80 %, for resistance (a) and susceptibility (b) to mastitis, resistance to bacterial mastitis (c), subclinical mastitis (d) and clinical mastitis (e) in a Mexican Braunvieh cattle population. PC1 = percentage of variability explained by dimension one. PC2 = percentage of variability explained by dimension two.



**Figure 7.** Biplots: marker's contribution and 10% most contributing individuals for resistance (a) and susceptibility (b) to mastitis, resistance to bacterial mastitis (c), subclinical mastitis (d) and clinical mastitis (e) in a Mexican Braunvieh cattle population. PC1 = percentage of variability explained by dimension one. PC2 = percentage of variability explained by dimension two. Note: the names of each locus by trait are indicated in Tables 1 to 5.

Oddly enough, for RM, the highly inbred animal 16 is among the top 10% of the most contributory animals (Figure 7). This can probably be explained by its position in the biplot, which means it is most associated with gene TBC1D8, which contributed little in the analysis. The same situation is true for SM, animal 143, and the marker for the gene ITPK1.

For SM, the markers for genes ANKRD33B, CTNND2, GRIA3, and SIRT1 contributed the most to the PCA, Figure 6. The pairs of genes CTNND2 with SIRT1, and ANKRD33B

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with GRIA3 had a positive relationship (Figure 7) even though they are in distant chromosomes. This may be a particular feature of the genotypic profile for mastitis in this Braunvieh population.

The markers of genes IL1A and ITGB3 for RBM were in a relationship similar to that of the genes ARHGAP10 and BDH2 for RM, Figure 6. They had a negative relationship according to the biplot, Figure 7. Similarly, they are in different chromosomes, even though both directly influence the function of the immune system [19–20].

The marker BovineHD0300029925 (10) of HIVEP3, Figures 6 and 7, contributed the most to the PCA for RSM. This SNP is positively related to another two intragenic HIVEP3 markers: BovineHD0300029904 (9) and BovineHD0300029955 (11). For RCM, most of the markers were highly contributive, i.e., the SNPs for the genes CXCL8, SEL1L, and STAT4, associated directly with the immune system [47].

#### 4. Conclusions

The population studied presented high genetic diversity for resistance and susceptibility to mastitis, even higher than that of the original Braunvieh and Brown Swiss. Although 13.3% were highly inbred animals (>50%), there were highly heterozygous groups in terms of the traits studied, a favorable indicator of the presence of diversity. Even though, the homozygous genotypes for immunologic genes (group CXCL) might be favorable for traits such as resistance to subclinical mastitis and resistance to clinical mastitis.

The K-means algorithm and the Principal Components Analysis are good statistical tools to visualize the groups of animals with different genotypic profiles. The Principal Components Analysis was useful to detect relationships and contributions of markers for each trait. The results of our research give evidence of the genetic potential of the Mexican Braunvieh population for improvement of mastitis-related traits in the tropic.

**Author Contributions:** Conceptualization, A.R.F. and M.Z.T.C.; methodology, M.Z.T.C.; software, P.P.R.; validation, P.P.R.; formal analysis, M.Z.T.C. and R.S.R.; investigation, A.R.F. and M.Z.T.C.; resources, A.R.F.; data curation, A.R.F.; writing—original draft preparation, M.Z.T.C. and R.S.R.; writing—review and editing, A.R.F. and P.P.R.; visualization, A.R.F.; supervision, A.R.F. and P.P.R.; project administration, A.R.F.; funding acquisition, A.R.F. All authors have read and agreed to the published version of the manuscript.

**Funding:** This research was partially funded by Universidad Autónoma Chapingo, grant number DGIP-166701012.

**Institutional Review Board Statement:** Ethical review and approval were waived for this study because the producers obtained the hair samples for genotyping at their farms following their procedures.

**Informed Consent Statement:** Not applicable.

**Data Availability Statement:** Upon request to the author for correspondence.

**Acknowledgments:** Authors thank the Consejo Nacional de Ciencia y Tecnología, Mexico, for the financial support to the first author during her Master of Science studies. The authors also thank the Asociación Mexicana de Criadores de Ganado Suizo de Registro for allowing the use of their databases and the breeders for their kind cooperation in this study.

**Conflicts of Interest:** The authors declare no conflict of interest.

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## **CAPÍTULO 6. CONCLUSIONES GENERALES**

Los capítulos 3 a 5 muestran evidencia de la alta diversidad genética en bovinos Suizo Europeo para características complejas, sinónimo de su versatilidad genética sobresaliente en comparación con sus razas emparentadas Braunvieh original y Pardo Suizo; debido a que esta población se ha desarrollado por más de un siglo bajo condiciones tropicales. La diversidad genética se midió mediante la heterocigosidad observada, coeficiente de consanguinidad, frecuencia de alelos favorables y estadística multivariada.

En el capítulo 3 se encontraron frecuencias mayores a las de otras razas para genotipos asociados con mayor suavidad, mayor marmoleo, menor cantidad de ácidos grasos saturados, y menor fuerza de corte (CAPN1 y DGAT2). Existen similitudes con las frecuencias de la mayoría de las razas Europeas para los genes CAPN3 y TG; mientras que, para los genes DGAT1 y ANK1 las frecuencias de los genotipos deseados son inferiores. Un marcador de DGAT1 y el marcador de MADH3 resultaron monomórficos.

En el estudio para características reproductivas (capítulo 4), los valores de  $H_o$  cercanos a 0.5, sugieren un incremento en su versatilidad debido al ambiente tropical, dado que el valor difiere al de los ancestros directos de la raza y coincide con los de razas adaptadas al trópico. Además, el gen DSC2 fue el más relevante en este estudio ya que explicó variabilidad en los grupos para todas las características reproductivas estudiadas.

Por último, en el capítulo 5, se encontró que, para resistencia y susceptibilidad a mastitis, los perfiles genotípicos mostraron alta diversidad genética entre grupos, donde resistencia a mastitis clínica tuvo la menor presencia de genotipos heterocigóticos. Aunque el porcentaje de animales altamente consanguíneos (>50%) es de hasta 13.3%, existen grupos altamente heterocigóticos para las características estudiadas, indicador favorable de la presencia de diversidad.

En general, el uso de estadística multivariada fue clave para encontrar a los grupos de animales con perfiles genotípicos similares y el número de animales heterocigóticos por característica, lo anterior fue útil para determinar el grado de diversidad genética de la población para las diferentes características estudiadas. Los resultados de estas investigaciones evidencian el potencial genético de la población Suizo Europeo mexicana, para mejorar características relacionadas con calidad de la carne, reproducción y resistencia a mastitis.